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Antioxidant properties of different products and additives in white wine

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Abstract: Different winemaking products (ascorbic acid, glutathione, yeast lees and a yeast autolysate) were tested in comparison with sulfur dioxide, concerning radical scavenging activity (measured by DPPH* assay), oxygen consumption capacity and ability to reduce wine color and predisposition to browning. Trials were performed in white wines and model solution. SO2 was the most active in reducing wine color development. Fresh lees and ascorbic acid were very effective in oxygen and free radical scavenging, but they both induced browning during wine storage, the former, by releasing phenolic compounds. Glutathione was also able to scavenge DPPH* in wine, but less effective against oxygen, and it induced browning during storage. Surprisingly, the yeast derivative preparation was the treatment that behave more similarly to sulfiting; it was very active in scavenging DPPH*, and, even without modifying oxygen consumption rate, it protected quite well wine color and phenolics over a eight months storage time.

Cover Letter

Sulfur dioxide is considered one of the most important enological additives, but it is also toxic and allergenic. In Europe, it is mandatory to label wines with the indication "contains sulfites", if total sulfur dioxide is higher than 10 mg/L; this has determined an increasing interest of the consumers towards the health-related aspects connected with wine consumption. For this reason, the reduction of the use of sulfur dioxide is considered one of the key objectives of modern winemaking and it is strictly connected with the priorities established by the "International Organization of Vine and Wine" (O.I.V.). Current knowledge and technologies allow to postpone sulfiting at the end of alcoholic and malolactic fermentation, with a significant reduction of the overall sulfite levels without jeopardizing wine quality. Contrary, it is more difficult to replace SO_2 after the fermentations, because none of the products currently available for complementing the activity of this additive during wine storage and ageing, is as effective as sulfites themselves. Some of these complementary substances, such as ascorbic acid, have been extensively studied, while some other (e.g. glutathione) were less investigated; anyway, maybe paradoxically, the most of the studies available have been published on technical journals, and the most of the few scientific publications available are focused just on one or two alternatives, sometimes without a direct comparison with sulfur dioxide; moreover, the trials have been generally performed in model solution and only few evidences have been collected on wines. This paper aims to investigate the radical scavenging activity and the oxygen consumption capacity of different enological products and additives (ascorbic acid, glutathione, yeast lees and a yeast derivative preparation), in comparison with SO₂, also considering their effect on wine color and predisposition to browning. Trials were performed in model solution and in different wine typologies. We consider these results just as a preliminary investigation, but, as far as we know, this is one of the most comprehensive, direct comparisons, between sulfur dioxide and other additives, reporting effects on real wines. Moreover, surprisingly, the yeast derivative preparation used, was the additive that behave more similarly to sulfur dioxide, demonstrating its ability in protecting wine color over a 8 months storage time. As far as we know, this is the first time this kind of products has been directly compared with sulfites concerning their antioxidant characteristics.

1 Antioxidant properties of different enological products

2 and additives in comparison with sulfur dioxide

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14 Abbreviated running title:

15 Antioxidant capacity of enological products compared with sulfites

16 Abstract

17 Different winemaking products (ascorbic acid, glutathione, yeast lees and a yeast autolysate) 18 were tested in comparison with sulfur dioxide, concerning radical scavenging activity (measured by DPPH• assay), oxygen consumption capacity and ability to reduce wine color 19 20 and predisposition to browning. Trials were performed in white wines and model solution. 21 SO₂ was the most active in reducing wine color development. Fresh lees and ascorbic acid 22 were very effective in oxygen and free radical scavenging, but they both induced browning 23 during wine storage, the former, by releasing phenolic compounds. Glutathione was also able to scavenge DPPH• in wine, but less effective against oxygen, and it induced browning during 24 25 storage. Surprisingly, the yeast derivative preparation was the treatment that behave more 26 similarly to sulfiting; it was very active in scavenging DPPH•, and, even without modifying oxygen consumption rate, it protected quite well wine color and phenolics over a eight 27 28 months storage time.

29

30 KEYWORDS: sulfur dioxide; yeast derivatives; ascorbic acid; glutathione; yeast lees; 31 wine

33 1 Introduction

34 Despite the mechanisms involved in wine oxidation have been extensively reviewed 35 (Singleton, 1987; du Toit, Marais, Pretorius, & du Toit, 2006; Waterhouse & Laurie, 2006; 36 Oliveira, Ferreira, De Freitas, & Silva, 2011), the protection of wine against oxidative 37 spoilage remains one of the main goals of modern winemaking, becoming particularly critical 38 when low levels of sulfur dioxide are used. The chemistry of this additive in wine has been 39 recently re-written by Danilewicz (2007; 2011) and Danilewicz, Seccombe, & Whelan 40 (2008): they clearly demonstrated that SO_2 does not react directly with oxygen, as previously thought (Ribéreau-Gayon, Dubourdieu, Doneche, & Lonvaud, 2006), but, in presence of 41 42 metal ions, it is able to scavenge hydrogen peroxide and the quinones formed from the 43 oxidation of polyphenols (Danilewicz et al., 2008). Due to the toxicity and allergenic potential 44 of sulfites, different compounds have been proposed for reducing their final concentration in 45 wine, even if, none of them is likewise effective in protecting wine against oxidations.

46 Ascorbic acid (ASC) is the most known among these products; it is able to scavenge hydroxyl 47 radicals (Bradshaw, Barril, Clark, Prenzler, & Scollary, 2011) and quinones (Waterhouse & Laurie, 2006; Bradshaw et al., 2011), but its metal catalyzed oxidation produces hydrogen 48 49 peroxide (Zoecklein, Fugelsang, Gump, & Nury, 1995; Moreaux, Birlouez-Aragon, & 50 Ducauze, 1996; Ribéreau-Gayon et al., 2006; Bradshaw et al., 2011) and this may trigger 51 browning reactions if sulfites are not present (Bradshaw, Prenzler, & Scollary, 2001; Bradshaw, Cheynier, Scollary, & Prenzler, 2003). The capacity of ascorbic acid to act both as 52 antioxidant and free-radical initiator is known as "crossover effect" (Buettner & Jurkiewicz, 53 54 1996; Bradshaw et al., 2001; Bradshaw et al., 2003) and explains the reason why ASC is normally used in wine in combination with sulfites (Zoecklein et al., 1995; Ribéreau-Gayon et 55 56 al., 2006; Bradshaw, et al., 2011).

57 Another traditional system to protect wine against oxidations is the use of yeast lees (Pérez-Serradilla & Luque de Castro, 2008). Fresh lees have a high oxygen consuming capacity 58 59 (Fornairon-Bonnefond & Salmon, 2003), due to the presence of yeast membrane lipids and 60 sterols (Salmon, Fornairon-Bonnefond, Mazauric, & Moutounet, 2000; Fornairon-Bonnefond 61 & Salmon, 2003); adsorbed polyphenols (Gallardo-Chacón, Vichi, Urpí, López-Tamames, & 62 Buxaderas, 2010), thiol groups of cell wall proteins (Jaehrig, Rohn, Kroh, Fleischer, & Kurz, 2007); Gallardo-Chacón et al., 2010) and β -glucans from yeast cell walls (Jaehrig et al., 2007) 63 64 also contributes to their antioxidant properties. However, ageing on the lees can modify wine 65 sensory characters, and for this reason it is not suitable for all the wine typologies; moreover, lees alone do not protect wine against microbial pollution and sulfiting is always required. 66

The possibility to use glutathione (GSH) as wine antioxidant has been considered since the role of this tripeptide in preventing must browning has been highlighted (Singleton, Salgues, Zaya, & Trousdale, 1985); nevertheless, very few studies are currently available concerning GSH addition in wine. It has been reported that glutathione supplementation could have positive effects on wine color and aroma (Dubourdieu & Lavigne-Cruege, 2003; Papadopoulou & Roussis, 2008), but high amounts of GSH in oxidative conditions can lead to color formation (Sonni, Clark, Prenzler, Riponi, & Scollary, 2011).

GSH can be supplemented also in form of yeast derivatives (YD): the ability of "glutathioneenriched" inactive dry yeast preparations (IDY) in reducing the loss of volatile compounds during wine storage has been reported by Andújar-Ortiz, Rodríguez-Bencomo, Moreno-Arribas, Martin-Alvarez, & Pozo-Bayon (2010): they hypothesized that this may be due to the antioxidant capacity of GSH, but also other components of the IDY preparation might be involved (Andújar-Ortiz et al., 2010).

The opportunity to reduce sulfur dioxide, by these alternative tools, is arousing more and more interest, among winemakers. Nevertheless, despite the amount of works reporting the antioxidant effects of these substances, the most of the papers regards model solutions and 4 moreover, it is currently difficult to foresee in which extent it is possible to replace sulfites
with each of these alternatives, preserving wine quality, because of the lack of scientificallybased direct comparisons, among their effects and those of sulfur dioxide.

86 For this reason, the aim of this work was to carry out a preliminary investigation on the 87 radical scavenging activity (measured by DPPH• assay) and the oxygen consumption capacity 88 of different enological products and additives in comparison with SO₂. Ascorbic acid 89 (considered as reference standard), glutathione, yeast lees and a self-prepared yeast autolysate 90 were tested. Trials were performed in model solution and in different wine typologies. 91 Concerning oxygen consumption trials, wines were finally subjected to fast 92 spectrophotometric measurements, for assessing the effect of the different antioxidants on 93 color, total phenolics and predisposition to browning.

94 **2** Materials and Methods

95 2.1 Chemicals

96 Tartaric acid, sodium hydroxide, ethanol (96 % v/v), ACS grade hydrochloric acid (37 %),
97 hydrogen peroxide (30 % w/w), sodium acetate and potassium metabisulfite were from Carlo
98 Erba Reagents (Milan, Italy); ascorbic acid, glutathione, 1,1-diphenyl-2-picryl-hydrazyl free
99 radical (DPPH•) and HPLC grade methanol were purchased from Sigma-Aldrich (St. Louis,
100 MO, USA).

101 2.2 Yeast lees and yeast derivatives (YD)

Fresh lees were supplied by Viticoltori Friulani "La Delizia" (Casarsa della Delizia, PN, Italy) and they were obtained after alcoholic fermentation and racking of a white table wine. The inactive dry yeast preparation (YD) used for the trials was a thermally produced yeast autolysate, prepared as reported elsewhere (Comuzzo, Tat, Liessi, Brotto, Battistutta, & Zironi, 2012).

108 2.3.1 Sample preparation

Trials were performed in model solution and wine. The former was a model buffer prepared by dissolving 5 g/L (33 mM) of tartaric acid in a distilled water – ethanol mixture (12 % v/v); the pH was set at 3.20 by adding 4 M sodium hydroxide. Ascorbic acid (50 mg/L), glutathione (50 and 500 mg/L), potassium metabisulfite (100 and 1000 mg/L, corresponding respectively to 50 and 500 mg/L of sulfur dioxide), yeast lees (2.5 % v/v) and the YD preparation (2.5 % w/v) were added and the samples were immediately analyzed by DPPH• assay, as reported below.

The wine was a white table wine from harvest 2010, supplied by Viticoltori Friulani "La Delizia" (Casarsa della Delizia, PN, Italy); additives, lees and YD preparation were added in the same amounts reported above for wine-like solution; in addition a Control sample (untreated wine) was also included in the experimental design. Control wine and treated samples were subjected to DPPH• assay as reported below. All the experiments were carried out in three repetitions, for both wines and model solutions.

122 2.3.2 DPPH• assay

123 DPPH• assay was performed by a modification of the methods reported by Brand-Williams, Cuvelier, & Berset (1995) and Gallardo-Chacón et al. (2010), using a UV-vis 124 spectrophotometer (model V-530, Jasco Co. Ltd., Tokyo, Japan). A 6×10^{-5} M DPPH• 125 126 solution was prepared fresh daily, in a 60:40 mixture of methanol : acetate buffer (0.1 M 127 sodium acetate, buffered at pH 4.50 with 6 M hydrochloric acid). 3 mL of this stock solution 128 were introduced in a 10 mm optical path length glass cuvette (Hellma Analytics, Mülheim, 129 Germany) and 100 µL of the wine samples or fresh prepared antioxidant model solutions were 130 added; DPPH• discoloration was followed at 515 nm during 10 min, reading the absorbance against methanol: acetate buffer. Results were expressed as the percent diminution of the original absorbance [Δ Abs 515 nm (%)].

For the samples treated with yeast lees and YD preparation, where insoluble particles were present, the reaction with DPPH• has been carried out as suggested by Gallardo-Chacón et al. (2010): 3 mL of DPPH• and 100 μ L of sample were introduced in a test tube; after 10 min, the reaction mixture was filtered on a 0,80 μ m nylon membrane and immediately subjected to spectrophotometric measurement. The initial value of the absorbance was read by adding 100 μ L of methanol: acetate buffer, to 3 mL of DPPH• stock solution.

prepared, performing the DPPH• assay on the model buffer alone (tartaric acid in hydroalcoholic solution 12 % v/v, pH 3.2); the percent values measured for the Δ Abs 515 nm were used to correct the analytical results.

143 2.4 Oxygen consumption capacity

144 2.4.1 Equipment

The system used for oxygen measurements was an OxySense[®] fluorimeter (OxySense Inc., 145 Dallas, TX, USA); O2xyDot® oxygen sensitive sensors (OxySense Inc., Dallas, TX, USA), 146 147 were glued, by a specific silicon based oxygen permeable adhesive (OxySense Inc.), to the 148 inner surface of each of the 750 mL colorless glass bottles used for the experiments. When O2xyDot[®] sensors are illuminated by a pulsed blue light, they emits a red fluorescent light, 149 that is monitored by OxySense[®] fluorimeter. Dynamic quenching by oxygen molecules 150 determines a decrease of the O2xyDot[®] fluorescence lifetime, that is proportional to the 151 152 oxygen concentration in the bottles; the temperature is measured simultaneously, by an 153 infrared sensor positioned in the reader pen (Li, Ashcraft, Freeman, Stewart, Jank, & Clark, 154 2008).

155 2.4.2 Sample preparation

156 Two different white wines were used in two different sets of experiments. In the first one, a 157 base wine for Prosecco D.O.C.G. Conegliano Valdobbiadene (harvest 2012), supplied by 158 Cantina Produttori Valdobbiadene "Val d'Oca" (San Giovanni di Valdobbiadene, TV, Italy), 159 was used; free sulfur dioxide was 13 mg/L and alcoholic strength 10.50 % v/v. The wine 160 coming from a 20 L bulk, was shaken until oxygen saturation was reached and then transferred into the 750 mL bottles where the O2xyDot[®] sensors were glued (see Section 161 162 2.4.1). The average oxygen concentration measured at filling was 8.25 ± 0.19 mg/L and the 163 temperature of the samples was 19.0 °C. Ascorbic acid (50 mg/L), glutathione (50 mg/L), 164 potassium metabisulfite (100 mg/L, corresponding to 50 mg/L of sulfur dioxide), yeast lees 165 (2.0 % v/v) and YD preparation (500 mg/L) were added to the bottles; all the treatments were 166 replicated three times. Nitrogen was blown in the headspace of the samples and bottles were 167 immediately sealed with crown cap closures. A control sample (untreated wine) without any 168 antioxidant addition was also prepared in three repetitions.

169 A blend of Chardonnay, Sauvignon and Picolit from harvest 2007 (Bastianich Winery, 170 Cividale del Friuli, UD, Italy) was used for the second set of trials; free sulfur dioxide was 5 171 mg/L and alcoholic strength 13.50 % v/v. Wine was saturated with oxygen (as reported above) and then transferred in the 750 mL bottles with O2xyDot[®] sensor; the average oxygen 172 173 concentration at filling was 7.29 ± 0.14 mg/L and the temperature of the samples 21.0 °C. 174 Potassium metabisulfite, ascorbic acid, glutathione, YD preparation and fresh lees were added 175 to the bottles in the same amounts reported above; a control sample (untreated wine) without 176 any antioxidant addition was also prepared and three repetitions were setup for each 177 experiment. As reported for the Prosecco base wine, nitrogen was blown in the headspace of 178 the samples and bottles were immediately crown capped. For both wine typologies, oxygen 179 concentration into the bottles was measured daily and wines were stored at 20 °C during the 180 whole time of the measurements.

181 2.4.3 Spectrophotometric measures and browning assay

All the wines were analyzed 15 days after bottling; in addition, for the blended wine, analyses were repeated after 8 months. Wine color and total phenolics were assessed by measuring the absorbance of the samples at 420 and 280 nm respectively, using 10 mm optical path length quartz cuvettes (Hellma Analytics, Mülheim, Germany); readings were performed against distilled water. Concerning the UV measures, wine samples were previously diluted ten times and total phenolic index (TPI) was calculated multiplying by 10 the absorbance measured at 280 nm.

The predisposition of wines towards browning was determined by slightly modifying the so called POM-test, a browning test reported by Müller-Späth (1992); briefly, 5 mL of wine were heated at 60 °C for one hour, after addition of 25 μ L of a 3 % hydrogen peroxide solution; the browning produced was estimated on the basis of the percent increase of the absorbance at 420 nm. All the analyses were carried out by using a UV-vis spectrophotometer, model V-530 (Jasco Co. Ltd., Tokyo, Japan).

195 2.5 Statistical analysis

As concerns DPPH• trials, One Way ANOVA was carried out on the percent diminutions of absorbance, measured during the assay [Δ Abs 515 nm (%)]; means and standard deviations (SD) were calculated and significant differences were evaluated by Tukey HSD test, at p < 0.05. Variances were homogeneous according to Brown-Forsythe test.

200 To assess differences in oxygen consumption rates, Factorial ANOVA was used on the 201 oxygen concentrations measured for the different treatments; means and standard errors (SE)

- 202 were calculated, and significant differences were assessed by Tukey HSD test (p < 0.05).
- 203 Variances were homogeneous according to Cochran C, Hartley F-max and Bartlett test.
- 204 Finally, as regards spectrophotometric measurements (Abs 280 nm, 420 nm and POM-test),
- 205 One Way ANOVA and Tukey HSD test were carried out as reported for DPPH• assay;

results were considered significant at p < 0.05. All the analyses were carried out by using the

207 software Statistica for Windows, version 8.0 (StatSoft, Inc., Tulsa, OK, USA).

208 **3 Results and Discussion**

209 3.1 Evaluation of radical scavenging activity

The results of DPPH• assay are reported in Fig. 1. Concerning model solutions (Fig. 1a), the highest radical scavenging activity was detected for yeast lees, followed by YD and the highest sulfur dioxide addition (500 mg/L). At lower amounts, such as those normally used in winemaking (50 mg/L), SO₂ was less effective in bleaching the free radical and the values reported were more similar (even if statistically lower) to those measured for the same dosage of ascorbic acid (reference standard).

Despite the reaction of sulfite (SO_3^{2-}) and bisulfite ion (HSO_3^{-}) with free radicals was 216 217 described (Brandt & van Eldik, 1995; Neta & Huie, 1985), very few publications report data 218 about their radical scavenging capacity in wine; moreover, the information available are 219 generally not in agreement with the dataset presented here. In a paper published in the late 220 1990s, Manzocco, Mastrocola, & Nicoli (1998) found that the addition of sulfur dioxide (250 221 mg/L) to a model system simulating wine (water - ethanol 12 % v/v), did not affect the chain-222 breaking capacity of the samples towards DPPH•; the different results they obtained respect to 223 the present study might be related to the different pH and ionic strength of the solvents used 224 in the two experiments, for both sample preparation (hydroalcoholic solution vs. 225 hydroalcoholic buffer, pH 3.20) and DPPH• assay (pure methanol vs. methanol : acetate 226 buffer pH 4.50). pH might have affected the ratio between the different forms of sulfur(IV) 227 oxides in aqueous solution and it is known that they have a different reactivity towards free 228 radicals (Neta & Huie, 1985). Moreover, also the different ionic strength of the two model 229 solutions might have a non-negligible role: in fact, as reported by Brandt & van Eldik

230 (1995), the reaction rate between sulfur(IV) oxides and radicals increases with the increase of 231 the ionic strength. Anyway, it is important to underline that the hydroalcoholic tartaric buffer 232 we used for dissolving or suspending the different products tested, gave a very poor effect in 233 bleaching DPPH• solution ($\Delta Abs 515 \text{ nm} = -3.4 \%$).

234 Also the results of Vivas and colleagues (Vivas, Saint-Cricq de Gaulejac, & Glories, 1997) 235 are in disagreement with the radical scavenging capacity of sulfur dioxide, reported in Fig. 1a. Studying the influence of SO_2 and ascorbic acid on the scavenging effect of tannins, they 236 237 wrote that, at usual enological concentrations, sulfites do not have a scavenging effect on 238 superoxide anion. Nevertheless, according to literature, superoxide radical reacts very slowly with sulfite (SO₃²⁻) and the rate constant of the reaction is very low (82 $M^{-1} s^{-1}$) (Neta & Huie. 239 240 1985).

241 As observed for SO₂, also glutathione activity in model wine depends on the concentration. 242 GSH levels in wine range from non-detectable values to 70 mg/L (Kritzinger, Bauer, & du 243 Toit, 2013a), with a relatively high variability; at a concentration close to these amounts (50 mg/L), glutathione demonstrated the lowest ability in bleaching DPPH• and a dosage ten 244 245 times higher (500 mg/L) was needed to significantly overtake the effect of the reference 246 sample (ASC 50 mg/L); nevertheless, regardless of the amounts added, GSH activity was 247 always significantly lower than that measured for the same addition of sulfur dioxide.

248 This last considerations is not in full agreement with the data published by Vivas, Vivas de 249 Gaulejac, & Nonier (2001); they measured the scavenging activity towards superoxide anion 250 of different antioxidants in model solution, in a range of concentration from 0.1 to 1 g/L; they 251 also found that ascorbic acid was more effective than an equal amount of sulfur dioxide and 252 glutathione, but in their experiment, the scavenging activities observed for the latter additives 253 were very similar.

254 Speaking from the chemical point of view, the effects reported for such molecules in Fig. 1a seem connected quite well with their molar concentration. Glutathione molar concentration 255 11

in the sample at 50 mg/L, the one with the lowest radical scavenging activity, corresponds to approx. 0.16 mM. Ascorbic acid and sulfur dioxide at 50 mg/L, were 0.28 and 0.78 mM respectively and determined a greater percentage of discoloration; the higher efficiency of the former, despite its lower molar concentration, confirms the higher capacity of ASC to scavenge free radicals (Vivas et al., 2001). Finally, GSH at 500 mg/L (1.6 mM) and the same quantity of SO₂ (7.8 mM) showed an increasing activity in DPPH• bleaching.

262 On the basis of this trend, a non-negligible part of the effects observable when comparing 263 different antioxidants in winemaking conditions (therefore, in the amounts normally used and 264 speaking in terms of mg/L) should be attributed not only to the chemical characteristics of 265 each antioxidant molecule in itself, but also to its molar concentration. This is certainly a key 266 point in evaluating antioxidant additives in comparison with sulfur dioxide; in fact, due to its 267 average concentration in wine (0-40 mg/L as free SO₂) and to the lower molecular weight 268 respect to the most suggested alternatives (ACS or GSH), the molar concentration of free SO₂ 269 (considering both the molecular fraction and bisulfite ion) is normally higher, respect to that 270 of these latter molecules. For instance, 240 mg/L of GSH and 137 mg/L of ascorbic acid 271 would be required for obtaining a 0.78 mM solution, the same molar concentration of sulfur 272 dioxide in the sample at 50 mg/L; these amounts are more than a half of the European legal 273 limit established for ASC (250 mg/L, according to the Regulation (EC) No 606/2009) and far 274 from the normal quantity of GSH naturally detectable in wine (Dubourdieu & Lavigne-275 Cruege, 2003; Kritzinger et al., 2013a; Fracassetti, Lawrence, Tredoux, Tirelli, Nieuwoudt, & 276 du Toit, 2011; Sonni et al., 2011).

Another interesting observation can be made, comparing the effects reported in Fig. 1a for the yeast autolysate (YD) and the two samples supplemented with glutathione. As reported by Kritzinger and colleagues (Kritzinger et al., 2013a), commercial YDs claim to preserve wine aroma, delaying the development of browning and oxidized notes; reduced GSH is generally considered the YD's most active component from this point of view (Pozo-Bayón,

282 Andújar-Ortiz, & Moreno-Arribas, 2009; Andújar-Ortiz et al., 2010; Kritzinger et al., 2013a). 283 According to Tirelli, Fracassetti, & De Noni (2010), glutathione content in yeast autolysates 284 ranges from 1 to 14 mg/g (0.33-4.60 mmol/100 g), a value close to the 0.1-1 % (1-10 mg/g) of 285 the Saccharomyces cerevisiae dry cell weight (Bachhawat et al., 2009). These quantities also 286 agree with the information available about the ability of such products to release the tripeptide 287 in model solution: Andújar-Ortiz and co-workers (Andújar-Ortiz, Pozo-Bayón, Moreno-288 Arribas, Martin-Alvarez, & Rodríguez-Bencomo, 2012) detected 1-2 mg/L of reduced GSH, 289 after addition of 0.3 g/L of a glutathione-enriched inactive dry yeast (GSH-IDY); these data 290 were confirmed by Kritzinger and colleagues (Kritzinger, Stander, & Du Toit, 2013b), who 291 detected a glutathione release between 1.45 and 2.53 mg/L, after the same supplementation 292 (0.3 g/L) with five different GSH-IDY preparations. On the basis of these quantities, we could 293 roughly quantify in 3-8 mg the average amount of free glutathione released by 1 g of YD 294 preparation.

In the present study, we made a yeast autolysate addition of 25 g/L (2.5 % w/v), 50-100 times higher respect to the normal amounts used in winemaking (250-500 mg/L). According to the calculations above, this would be able to release approx. 75-200 mg/L of free GSH, so a lower quantity respect to the highest amount of glutathione we have added to the model wine (500 mg/L); nevertheless, the DPPH• discoloration promoted by YD in Fig. 1a is significantly higher respect to that determined by 500 mg/L of GSH.

301 On the basis of these results, we can hypothesize that, probably, glutathione was not the only 302 factor in determining the radical scavenging capacity of the yeast derivative, but something 303 else among YD components might be involved, with a non-negligible contribution, in the 304 effects observed. This hypothesis was also considered in the paper published by Andújar-305 Ortiz et al. (2010): reporting the effects of glutathione-enriched inactive dry yeast 306 preparations on the aroma of wines, besides the antioxidant capacity of GSH, they also 307 mentioned "the activation of different types of chemical reactions promoted by other308 components from the IDY preparations".

309 The most accredited among these "other components" might be proteins: Jaehrig et al. (2007) 310 consider proteins from yeast cell walls as one of the most active components from the anti-311 oxidative point of view, due to their aromatic side chains and thiol groups of cysteine 312 residues. In support of this postulate, Tirelli and colleagues (Tirelli et al., 2010) quantified the 313 amounts of reducing proteins with cysteine residues (RPC), in several YD products, in 314 comparison with the content of free glutathione (GSH) and free cysteine (Cys); RPC were 315 estimated at non-negligible concentrations in the four yeast autolysates evaluated (0.73-1.40 316 mmol in 100 g of product), with values close, or even higher, respect to that of reduced GSH. 317 Obviously, in the light of these few bibliographic evidences, our current dataset must be 318 considered only as a preliminary result, being still quite poor to allow a certain conclusion; 319 further investigations will be needed to confirm these hypotheses, as well as to better 320 elucidate the role of the different YD components and fractions in scavenging free radicals.

321 A final consideration about Fig. 1a is related to the comparison between the radical 322 scavenging activity of the YD preparation and that of yeast lees. The high amount of yeast 323 autolysate used in this study was selected to make it comparable with lees addition (2.5 % 324 v/v). Obviously, we can expect that fresh lees were less concentrated in yeast cell residues, 325 because of their higher humidity content; nevertheless, the anti-radical activity of the model 326 wines treated with lees was significantly higher respect to that observed for the samples 327 treated with the yeast derivative. This could be due to the presence of residual phenolic 328 substances in the lees, that could have contributed to DPPH• discoloration, but also to the 329 production process of yeast derivatives, which could have reduced the radical scavenging 330 capacity of yeast cell components. This last observation could confirm the conclusions of 331 Tirelli et al. (2010); in the study mentioned above, they also reported the levels of overall Cys (Cys + $2 \times$ Cystine), that is to say a parameter which also takes into account the thiol 332

333 groups in form of disulfide; this fraction was generally the most representative among those 334 estimated (free GSH, free Cys, RPC and overall Cys), with a big variation among the 335 analyzed YD formulations; for this reason, the authors hypothesized that the technologies 336 applied for the production of yeast derivatives are not suitable for preserving the thiol groups 337 of the proteins with cysteine residues, as well as the antioxidant capacity of these products.

338 When the radical scavenging activity was measured in a white wine, the results were different 339 respect to what observed in model solution (Fig. 1b). First of all, all the tested substances 340 significantly increased the DPPH• bleaching capacity of the wine itself (Control sample). 341 Lees was the most active product in promoting discoloration, but the differences among the 342 treatments seemed less intense than those observed in the model buffer. The concentration 343 affected in a lower extent the scavenging ability of the products: for example, the two levels 344 tested for sulfur dioxide gave, statistically, similar performances in terms of $\Delta Abs \%$ and also 345 the two dosages of glutathione were not as different as those reported in Fig. 1a.

346 These behaviors might be explained considering a sort of "matrix-related effect", connected to 347 the ability of the different antioxidants, to interact with other components present in wine. In 348 effects, the wine itself, demonstrated a non-negligible capacity in scavenging DPPH•; the 349 presence of the antioxidant products might have affected this capacity, not only by directly 350 reacting with the free radical, but also by interacting with certain wine compounds. For 351 instance, it is reported that ascorbic acid, glutathione and SO₂ can hamper oxidation chains by 352 regenerating phenolic species from o-quinones (Waterhouse & Laurie, 2006; Danilewicz, 353 2007; Bradshaw et al., 2011); phenolics are well known radical scavengers and this 354 regeneration could be connected just with the increased chain-breaking activity measured for 355 the treated samples.

356 It is also interesting to observe, that comparing the activity of GSH with those of ascorbic 357 acid and SO₂ (samples at 50 mg/L), the former is no more the less effective in promoting 358 DPPH• discoloration (as it was in model buffer): in Fig. 1b, at a low dosage, glutathione 15 behaves similarly to sulfur dioxide, being not statistically different respect to the performances of ascorbic acid. In addition, at 500 mg/L, despite the five-folds lower molar concentration and in opposition to what seen in wine-like solution, GSH demonstrated a higher efficacy, respect to sulfites.

363 Finally, a last consideration concerns the YD preparation; if compared just with glutathione 364 (500 mg/L), one can observe that, in wine, the differences between the two treatments are 365 basically nil. This might be explained considering the previously mentioned "matrix-related 366 effect", so that the supplementation with pure glutathione, in a real wine, seemed to be as 367 effective as the YD product, probably due to the capacity of the tripeptide to improve the 368 overall radical scavenging activity of the wine itself. This might be connected with the ability 369 of GSH to react with quinones, regenerating phenolic molecules (Waterhouse & Laurie, 370 2006), but further investigations shall be done to better clarify the role of glutathione and YDs 371 in hampering radical chains in both model solution and different wine typologies.

372 *3.2 Oxygen consumption capacity*

373 The ability of the different antioxidant products to affect oxygen consumption was 374 investigated in two different wines. The curves describing oxygen consumption vs. time, in a 375 young wine (harvest 2012), are reported in Fig. 2. As one can observe, the amount of oxygen 376 dissolved at saturation was completely depleted in the untreated wine (Control) in 192 hours 377 (8 days). The most active oxygen scavenging substances among the tested products were fresh 378 lees (2 % v/v) and ascorbic acid (50 mg/L). At the concentration used, pure glutathione (50 379 mg/L) and the yeast derivative preparation (500 mg/L) gave results similar to those observed in the Control samples. On the contrary, sulfur dioxide (50 mg/L) allowed to significantly 380 381 reduce oxygen consumption, with a final average level close to 3 mg/L. In Fig. 3, the 382 statistical confirmation of these behaviors is reported, on the basis of the results of Factorial 383 ANOVA. Concerning SO_2 , it is interesting to underline that the reduction of oxygen

384 depletion rate occurred in the last 48 hours of the monitoring period, while during the first six 385 days, sulfites did not basically affect the kinetic of oxygen consumption. This behavior seems 386 in contrast with that reported by Danilewicz and colleagues (Danilewicz et al., 2008): 387 studying the interaction of oxygen, sulfur dioxide and 4-methylcatechol in a model wine 388 containing iron and copper, they found that the rate of reaction of oxygen was accelerated by 389 SO_2 addition; the explanation they gave, is that sulfites can react with quinones accelerating 390 catechol autoxidation. They also confirmed this effect on a red wine where sulfites were 391 eliminated by adding hydrogen peroxide; this elimination significantly reduced the ability of 392 the wine to consume oxygen (Danilewicz et al., 2008).

393 On the other hand, a further confirmation of these results comes also from Fig. 4, where 394 oxygen consumption vs. time is reported for the aged wine (harvest 2007). As one can 395 observe, fresh lees and ascorbic acid were again the most active substances in scavenging 396 oxygen, but sulfur dioxide also increased the O_2 consumption rate; it was only a slight 397 acceleration, but according to the results of Factorial ANOVA, oxygen depletion was 398 significantly faster in the sulfited samples than in the Control wines (Fig. 5).

399 On the basis of these evidences, it is difficult to explain the reason why sulfites reduced the 400 oxygen consumption rate in the young wine (Fig. 2). This product was quite poor in phenolic 401 compounds (the average total phenolic index of the Control samples was 3.5) and had a 402 relatively low content of free SO_2 (see Section 2.4.2). If we consider that sulfites can 403 accelerate catechol autoxidation, it can be hypothesized that the addition of potassium 404 metabisulfite has actually accelerated the conversion of polyphenols into quinonic species. It 405 has been reported that, when "4-methylcatechol is oxidized in presence of SO₂, the 38 % 406 approximately of the quinone formed reacts with bisulfite to produce the sulfonic adduct and 407 the most of the remainder is reduced back to catechol" (Danilewicz et al., 2008); so, we 408 should expect a progressive reduction of the concentration of available polyphenols during the 409 storage time. Due to the low TPI of the wine, and due to the fact that, as already mentioned,

this reduction should have been faster in the sulfited samples, we can hypothesize that oxygen
consumption, was slowed down, in the last part of the curves of the latters (see Fig. 2), just
because of this faster consumption of reactive substances.

413 Another interesting observation can be done comparing the oxygen consumption curves 414 detected for the aged wine (Fig. 4) and the youngest one (Fig. 2); as one can observe, oxygen 415 disappearance was initially faster in the former, but the slope of the curves progressively 416 decreased, and after 216 hours the average level of residual oxygen (in the most of the 417 treatments) was still close to 1-1.5 mg/L; otherwise, the curves reported for the young wine 418 were less steep in their first part, but the complete oxygen consumption was obtained in less 419 than 200 hours (with the only exception of the sulfited samples). This different behavior is 420 probably related with the different phenolic content and "oxidative history" of the two wines. 421 The aged one had an average TPI of 11.6, while for the youngest one TPI was lower (only 422 3.5); the former was produced by prolonged barrel ageing, while the latter was stored in 423 stainless steel vats, so in a more protective environment with respect to oxygen contact.

424 So, the faster oxygen consumption, detected at the beginning of the monitoring period in the samples from harvest 2007, was probably due to their higher phenolic content, that might 425 426 have initially increased oxygen consumption capacity (Danilewicz et al., 2008); nevertheless, 427 the more intense contact that this wine had with oxygen during the whole production chain, 428 might have been the most important factor, responsible of the decrease of oxygen 429 consumption rate in the last part of the monitoring period. Contrary, in the youngest wine, 430 which had a lower TPI, but was probably subjected to a less intense aeration during 431 processing, oxygen depletion started slower, but it was completed in a shorter time, denoting 432 a higher capacity of such product to scavenge oxygen.

433 *3.3 Effect on wine color and predisposition to browning*

The effects of the different treatments on color and predisposition to browning of the young wine (harvest 2012) are reported in Table 1 (Section a). As one can observe, sulfiting allowed the lowest color development, followed by the treatment with YD preparation. GSH and ascorbic acid gave intermediate results, while the highest color formation was detected for Control samples and the wines treated with fresh lees.

The POM-test values confirm these behaviors: the highest levels mark out samples in which phenolic fraction is more preserved, while oxidative phenomena tend to reduce the POM-test index. It is interesting to underline that the lowest POM-tests were detected for the wines treated with GSH and ASC, while oxidizable phenolics were better protected by sulfur dioxide and YD preparation.

444 Concerning the aged wine (harvest 2007), the results were less evident (Table 1, Section b); 445 15 days after the treatments, only fresh lees gave a significant increase of the color, probably 446 due to their ability to release phenolic compounds (TPI significantly increased, according to 447 ANOVA, from a value of 11.6 ± 0.2 in the Control wines, to 15.0 ± 0.3 in the lees added 448 samples). Anyway, according to the POM-Test levels, after 15 days, potassium metabisulfite 449 and YD were the additives that allowed the best preservation of oxidizable phenolics; 450 concerning sulfites, this confirms the behaviors observed in Table 1a for the young wine, 451 while, in this case (Table 1b), the effects given by the yeast autolysate were less evident, and very close to those of glutathione and ascorbic acid. 452

The situation of the aged wine changed 8 months after the treatments (Table 1, Section b). Sulfites demonstrated their effectiveness in protecting wine against browning, allowing only a slight increase of the color during storage time. YD was the additive that behave more similarly to SO_2 : the color index (Abs 420 nm) of the samples treated with the autolysate preparation was significantly higher than those of the sulfited wines, but significantly lower than in all the other experiments, highlighting a certain ability of such kind of preparations 10 459 in protecting wine against color development, over a medium-long storage time. Finally, fresh460 lees and particularly ascorbic acid were the treatments which gave the most intense browning.

461 POM-test results confirm these considerations: SO₂ gave the highest index, followed by YD

462 and GSH, while the lowest values were detected just for ASC and fresh lees.

In conclusion, the products tested in this study confirmed to have different effects concerning
their antioxidant properties, and this reflects in a very different behavior towards the
modifications they can induce in wine.

466 Fresh lees were very effective in oxygen and free radical scavenging, but they increased wine 467 color by the release of phenolic compounds. Ascorbic acid confirmed its effectiveness in the 468 removal of oxygen and DPPH• free radical, but as reported elsewhere (Bradshaw et al., 2001), 469 it induced an intense browning during wine storage. Glutathione also demonstrated its ability 470 in scavenging DPPH• in wine; nevertheless, it showed a poor capacity in scavenging oxygen, 471 and its activity in protecting wine against color formation was lower than that given by 472 sulfiting (particularly after 8 months of storage). This confirms the data published by Sonni et 473 al. (2011), who reported that GSH can initially provide protection against oxidative 474 coloration, but eventually induced color formation.

475 Strictly speaking in terms of the possibility to replace sulfur dioxide, the analyses carried out 476 on wine color and predisposition to browning, highlighted that SO₂ remains the most 477 performing additive. Nevertheless, maybe surprisingly, the addition of the yeast derivative 478 preparation (YD), was the treatment that behave more similarly to sulfiting: YD was very 479 active in scavenging DPPH• free radical in both model solution and wine, and, even without 480 significantly affecting the ability of the wine to consume oxygen, it protected quite well color 481 and phenolics over a medium-length storage time (8 months). Due to the effects we have 482 observed for GSH, and the average levels reported for glutathione in inactive dry yeasts, we 483 are oriented to think that this ability of YDs might be not only connected with their capacity 484 to release the tripeptide in wine; the glutathione released might contribute, but we can

485 hypothesize that other components of these complex preparations might be involved in the486 effects observed, with a non-negligible role.

487 Obviously, the bottleneck in using yeast derivatives for partially replacing sulfites in bottled 488 wines is connected with the presence of insoluble particles in the commercial preparations, 489 but this does not jeopardize the possibility to use these products during the storage in stainless 490 steel containers. From this point of view, these specific results seem interesting, because such 491 kind of preparations might allow a significant reduction of sulfite levels during wine storage, 492 with the possibility of postponing sulfiting in the production steps immediately preceding 493 bottling. Further investigations will make it possible to confirm these behaviors, clarifying the 494 mechanisms through which YDs carry out their protective action, and allowing both the 495 optimization of their use in winemaking as antioxidant formulations and the development of 496 suitable technologies for producing specific preparations for this specific winemaking use.

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502 **References**

503 Andújar-Ortiz, I., Rodríguez-Bencomo, J.J., Moreno-Arribas, M.V., Martin-Alvarez, P.J.,

504 & Pozo-Bayón, M.A. (2010). Role of glutathione enriched inactive yeast preparations on

- the aroma of wines. In *Proceedings of* 33^{rd} World Congress of Vine and Wine 8^{th}
- 506 *General Assembly of the OIV* (pp. 154-161). Tiblisi, Georgia.
- 507
- Andújar-Ortiz, I., Pozo-Bayon, M.A., Moreno-Arribas, M.V., Martin-Alvarez, P.J., &
 Rodríguez-Bencomo, J.J. (2012). Reversed-phase high-performance liquid₂₁

| 510 | chromatography-fluorescence detection for the analysis of glutathione and its precursor γ - |
|-----|--|
| 511 | glutamyl cysteine in wines and model wines supplemented with oenological inactive dry |
| 512 | yeast preparations. Food Analytical Methods, 5, 154-161. |
| 513 | |
| 514 | Bachhawat, A.K., Ganguli, D., Kaur, J., Kasturia, N., Thakur, A., Kaur, H., Kumar, A., & |
| 515 | Yadav, A. (2009). Glutathione production in yeast. In T. Satyanarayana, & G. Kunze |
| 516 | (Eds.), Yeast biotechnology: diversity and applications (pp. 259-281). Berlin: Springer. |
| 517 | |
| 518 | Bradshaw, M.P., Barril, C., Clark, A.C., Prenzler, P.D., & Scollary, G.R. (2011). Ascorbic |
| 519 | acid: a review of its chemistry and reactivity in relation to a wine environment. Critical |
| 520 | Reviews in Food Science and Nutrition, 51, 479-498. |
| 521 | |
| 522 | Bradshaw, M.P., Cheynier, V., Scollary, G.R., & Prenzler, P.D. (2003). Defining the |
| 523 | ascorbic acid crossover from anti-oxidant to pro-oxidant in a model wine matrix |
| 524 | containing (+)-catechin. Journal of Agricultural and Food Chemistry, 51, 4126-4132. |
| 525 | |
| 526 | Bradshaw, M.P., Prenzler, P.D., & Scollary, G.R. (2001). Ascorbic acid-induced |
| 527 | browning of (+)-catechin in a model wine system. Journal of Agricultural and Food |
| 528 | Chemistry, 49, 934–939. |
| 529 | |
| 530 | Brand-Williams, W., Cuvelier, M., & Berset, C. (1995). Use of a free radical method to |
| 531 | evaluate antioxidant activity. Lebensmittel-Wissenschaft und-Technologie, 28, 25-30. |
| 532 | |
| 533 | Brandt, C., & van Eldik, R. (1995). Transition metal-catalyzed oxidation of sulfur(IV) |
| 534 | oxides. Atmospheric-relevant processes and mechanisms. Chemical Reviews, 95, 119-190. |
| 535 | |

| 536 | Buettner, G., & Jurkiewicz, B.A. (1996). Chemistry and biochemistry of ascorbic acid. In |
|-----|---|
| 537 | E. Cadenas, & L. Packer (Eds.), Handbook of Antioxidants (pp. 91-115). New York: |
| 538 | Marcel Dekker. |
| 539 | |
| 540 | Comuzzo, P., Tat, L., Liessi, A., Brotto, L., Battistutta, F., & Zironi, R. (2012). Effect of |
| 541 | different lysis treatments on the characteristics of yeast derivatives for winemaking. |
| 542 | Journal of Agricultural and Food Chemistry, 60, 3211-3222. |
| 543 | |
| 544 | Danilewicz, J.C. (2007). Interaction of sulfur dioxide, polyphenols, and oxygen in a wine- |
| 545 | model system: central role of iron and copper. American Journal of Enology and |
| 546 | <i>Viticulture</i> , <i>58</i> , <i>53-60</i> . |
| 547 | |
| 548 | Danilewicz, J.C. (2011). Mechanism of autoxidation of polyphenols and participation of |
| 549 | sulfite in wine: key role of iron. American Journal of Enology and Viticulture, 62, 319- |
| 550 | 328. |
| 551 | |
| 552 | Danilewicz, J.C., Seccombe, J.T., & Whelan, J. (2008). Mechanism of interaction of |
| 553 | polyphenols, oxygen, and sulfur dioxide in model wine and wine. American Journal of |
| 554 | Enology and Viticulture, 59, 128-136. |
| 555 | |
| 556 | Dubourdieu, D., & Lavigne-Cruege, V. (2003). The role of glutathione on the aromatic |
| 557 | evolution of dry white wine. Wine Internet Technical Journal, 17, 1-10 |
| 558 | (www.infowine.com). |
| 559 | |
| 560 | du Toit, W.J., Marais, J., Pretorius, I.S., & du Toit, M. (2006). Oxygen in must and wine: |
| 561 | a review. South African Journal of Enology and Viticulture, 27, 76-94. |

| 562 | |
|-----|--|
| 563 | Fornairon-Bonnefond, C., & Salmon, JM. (2003). Impact of oxygen consumption by |
| 564 | yeast lees on the autolysis phenomenon during simulation of wine aging on lees. Journal |
| 565 | of Agricultural and Food Chemistry, 51, 2584–2590. |
| 566 | |
| 567 | Fracassetti, D., Lawrence, N., Tredoux, A.G.J., Tirelli, A., Nieuwoudt, H.H., & du Toit, |
| 568 | W.J. (2011). Quantification of glutathione, catechin and caffeic acid in grape juice and |
| 569 | wine by a novel ultra-performance liquid chromatography method. Food Chemistry, 128, |
| 570 | 1136–1142. |
| 571 | |
| 572 | Gallardo-Chacón, J.J., Vichi, S., Urpí, P., López-Tamames, E., & Buxaderas, S. (2010). |
| 573 | Antioxidant activity of lees cell surface during sparkling wine sur lie aging. International |
| 574 | Journal of Food Microbiology, 143, 48-53. |
| 575 | |
| 576 | Jaehrig, S.C., Rohn, S., Kroh, L.W., Fleischer, LG., & Kurz, T. (2007). In vitro potential |
| 577 | antioxidant activity of $(1-3)$, $(1-6)$ - β -D-glucan and protein fractions from Saccharomyces |
| 578 | cerevisiae cell walls. Journal of Agricultural and Food Chemistry, 55, 4710–4716. |
| 579 | |
| 580 | Kritzinger, E.C., Bauer, F.F., & du Toit, W.J. (2013a). Role of glutathione in winemaking: |
| 581 | a review. J. Agric. Food Chem., 61, 269-277. |
| 582 | |
| 583 | Kritzinger, E.C., Stander, M.A., & Du Toit, W.J. (2013b). Assessment of glutathione |
| 584 | levels in model solution and grape ferments supplemented with glutathione-enriched |
| 585 | inactive dry yeast preparations using a novel UPLC-MS/MS method. Food additives & |
| 586 | contaminants. Part A, Chemistry, Analysis, Control, Exposure & Risk Assessment, 30, 80- |
| 587 | 92. |

| 588 | |
|-----|---|
| 589 | Li, H., Ashcraft, K., Freeman, B.D., Stewart, M.E., Jank, M.K., & Clark, T.R. (2008). |
| 590 | Non-invasive headspace measurement for characterizing oxygen-scavenging in polymers. |
| 591 | Polymer, 49, 4541–4545. |
| 592 | |
| 593 | Manzocco, M., Mastrocola, D., & Nicoli, M.C. (1998). Chain-breaking and oxygen |
| 594 | scavenging properties of wine as affected by some technological procedures. Food |
| 595 | Research International, 31, 673-678. |
| 596 | |
| 597 | Moreaux, V., Birlouez-Aragon, I., & Ducauze, C. (1996). Copper chelation by tryptophan |
| 598 | inhibits the copper-ascorbate oxidation of tryptophan. Redox Reports, 2, 191-197. |
| 599 | |
| 600 | Müller-Späth, H. (1992). Der POM-Test. Deutscher Weinbau, 23, 1099-1100. |
| 601 | |
| 602 | Neta, P., & Huie, R.E. (1985). Free-radical chemistry of sulfite. Environmental Health |
| 603 | Perspectives, 64, 209-217. |
| 604 | |
| 605 | Oliveira, C.M., Ferreira, A.C.S., De Freitas, V., & Silva, A.M.S. (2011). Oxidation |
| 606 | mechanisms occurring in wines. Food Research International, 44, 115-1126. |
| 607 | |
| 608 | Papadopoulou, D., & Roussis, I.G. (2008). Inhibition of the decrease of volatile esters and |
| 609 | terpenes during storage of a white wine and a model wine medium by glutathione and N- |
| 610 | acetylcysteine. International Journal of Food Science and Technology, 43, 1053–1057. |
| 611 | |
| 612 | Pérez-Serradilla, J.A., & Luque de Castro, M.D. (2008). Role of lees in wine production: a |
| 613 | review. Food Chemistry, 111, 447-456. |

| 614 | |
|-----|--|
| 615 | Pozo-Bayón, M.Á., Andújar-Ortiz, I., & Moreno-Arribas, M.V. (2009). Scientific |
| 616 | evidences beyond the application of inactive dry yeast preparations in winemaking. Food |
| 617 | Research International, 42, 754-761. |
| 618 | |
| 619 | Ribéreau-Gayon, P., Dubourdieu, D., Doneche, B., & Lonvaud, A. (2006). Handbook of |
| 620 | Enology. Vol. 1. The Microbiology of Wine and Vinifications. (2 nd ed.). New York: John |
| 621 | Wiley & Sons. |
| 622 | |
| 623 | Salmon, JM., Fornairon-Bonnefond, C., Mazauric, JP., & Moutounet, M. (2000). |
| 624 | Oxygen consumption by wine lees: impact on lees integrity during wine ageing. Food |
| 625 | Chemistry, 71, 519-528. |
| 626 | |
| 627 | Singleton, V.L. (1987). Oxygen with phenols and related reactions in musts, wines, and |
| 628 | model systems: observations and practical implications. American Journal of Enology and |
| 629 | Viticulture, 38, 69-77. |
| 630 | |
| 631 | Singleton, V.L., Salgues, M., Zaya, J., & Trousdale, E. (1985). Caftaric acid |
| 632 | disappearance and conversion to products of enzymic oxidation in grape must and wine. |
| 633 | American Journal of Enology and Viticulture, 36, 50-56. |
| 634 | |
| 635 | Sonni, F., Clark, A.C., Prenzler, P.D., Riponi, C., & Scollary, J.R. (2011). Antioxidant |
| 636 | action of glutathione and the ascorbic acid/glutathione pair in a model white wine. Journal |
| 637 | of Agricultural and Food Chemistry, 59, 3940–3949. |
| 638 | |

| 639 | Tirelli, A., Fracassetti, D., & De Noni, I. (2010). Determination of reduced cysteine in |
|-----|--|
| 640 | oenological cell wall fractions of Saccharomyces cerevisiae. Journal of Agricultural and |
| 641 | Food Chemistry, 58, 4565-4570. |
| 642 | |
| 643 | Vivas, N., Saint-Cricq de Gaulejac, N., & Glories, Y. (1997). Influence de SO ₂ et de |
| 644 | l'acide ascorbique sur l'activité antiradicalaire des tanins, mesurée sur l'anion superoxyde. |
| 645 | Application aux vins rouges. Vitis, 36, 91-96. |
| 646 | |
| 647 | Vivas, N., Vivas de Gaulejac, N., & Nonier, M.F. (2001). Scavenging and antioxidant |
| 648 | properties of different soluble compounds in hydroalcoholic media. Comparison with SO_2 |
| 649 | efficiency. Bulletin de l'O.I.V., 843-844, 331-346. |
| 650 | |
| 651 | Waterhouse, A.L., & Laurie, V.F. (2006). Oxidation of wine phenolics: a critical |
| 652 | evaluation and hypotheses. American Journal of Enology and Viticulture, 57, 306-313. |
| 653 | |
| 654 | Zoecklein, B.W., Fugelsang, K.C., Gump, B.H., & Nury, F.S. (1995). Sulfur Dioxide and |
| 655 | Ascorbic Acid. In, Wine Analysis and Production (pp. 178-191). New York: Chapman |
| 656 | and Hall. |

657 Figure Captions

Fig. 1. Radical scavenging activity of different enological additives and products in model solution (a) and in white table wine (b). Results of ANOVA analysis and Tukey HSD test; means and standard deviations of three repetitions are reported. Different letters mark significant differences at p < 0.05. See the text for abbreviations.

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Fig. 2. Oxygen consumption versus time, measured for the samples prepared in Prosecco base wine
(harvest 2012). Mean values of three repetitions are reported. Vertical bars represent standard
deviations. See the text for abbreviations.

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Fig. 3. Factorial ANOVA and Tukey HSD test, carried out on the oxygen concentrations measured for the curves in Fig. 2. Means and standard errors (SE) of three repetitions are reported; different letters mark significant differences at p < 0.05. See the text for abbreviations.

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Fig. 4. Oxygen consumption versus time, measured for the samples prepared in the blended wine
from harvest 2007 (Chardonnay, Sauvignon, Picolit). Mean values of three repetitions are reported.
Vertical bars represent standard deviations. See the text for abbreviations.

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Fig. 5. Factorial ANOVA and Tukey HSD test, carried out on the oxygen concentrations measured for the curves in Fig. 4. Means and standard errors (SE) of three repetitions are reported; different letters mark significant differences at p < 0.05. See the text for abbreviations.

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726 **Table 1**

727 Absorbance at 420 nm and POM-test values detected for the two wines used in the experiments; samples were analyzed 15 days after bottling, and

128 limitedly to the aged wine (harvest 2007), after 8 months. Means, standard deviations (SD) and the results of One Way ANOVA and Tukey HSD test are

reported; different letters represent significant differences at p < 0.05.

| (a) Young v | vine (harvest 2012) | | | (b) Aged wine (harve | est 20 | 07) | |
|-------------|----------------------|---|-------------|----------------------|--------|---------------------|--|
| Abs 420 nm | | | | Abs 420 nm | | | |
| Sample | 15 days Mean ± SD | | Sample | 15 days | | 8 months | |
| | | | | Mean ± SD | | Mean ± SD | |
| Control | 0.038 ± 0.001 | e | Control | 0.204 ± 0.002 | a | 0.251 ± 0.005 | |
| SO2 50 mg/L | $0.019~\pm~0.000$ | а | SO2 50 mg/L | $0.182 ~\pm~ 0.002$ | a | $0.200~\pm~0.000$ | |
| YD 500 mg/L | $0.024 ~\pm~ 0.000$ | b | YD 500 mg/L | $0.206~\pm~0.024$ | a | $0.214 ~\pm~ 0.009$ | |
| Lees (2 %) | $0.039 ~\pm~ 0.000$ | e | Lees (2 %) | $0.273 ~\pm~ 0.010$ | b | $0.280~\pm~0.005$ | |
| ASC 50 mg/L | $0.033 ~\pm~ 0.001$ | d | ASC 50 mg/L | $0.198~\pm~0.001$ | a | $0.288~\pm~0.001$ | |
| GSH 50 mg/L | $0.028 ~\pm~ 0.000$ | с | GSH 50 mg/L | $0.205 ~\pm~ 0.001$ | a | $0.258~\pm~0.005$ | |

| | POM-test | | |
|-------------|--------------|----|--|
| Sample | 15 days | | |
| | Mean ± SD | | |
| Control | 90 ± 10 | b | |
| SO2 50 mg/L | 167 ± 36 | cd | |
| YD 500 mg/L | 181 ± 6 | d | |
| Lees (2 %) | 111 ± 17 | bc | |
| ASC 50 mg/L | 20 ± 35 | a | |
| GSH 50 mg/L | 25 ± 2 | а | |

| | POM-test | | | | |
|-------------|-------------|----|------------|----|--|
| Sample | 15 days | | 8 months | | |
| | Mean ± SD | | Mean ± SD | | |
| Control | 35 ± 15 | а | 24 ± 2 | bc | |
| SO2 50 mg/L | 69 ± 5 | b | 45 ± 0 | e | |
| YD 500 mg/L | $43~\pm~14$ | ab | 29 ± 3 | d | |
| Lees (2 %) | 30 ± 9 | а | 21 ± 1 | ab | |
| ASC 50 mg/L | 42 ± 1 | а | 19 ± 0 | а | |
| GSH 50 mg/L | 42 ± 5 | а | 27 ± 1 | cd | |

Highlights

Sulfur dioxide was the most active additive in reducing wine color development Ascorbic acid was a good O_2 and radical scavenger but induced browning during storage Lees were good O_2 and radical scavengers but increased color by releasing phenolics Glutathione had radical scavenging ability in wine but increased color during storage Yeast derivative behave more similarly to SO_2 protecting wine color over 8 months