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1	Influence of lipoproteins at dry-off on metabolism of dairy cows during transition period and
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23	analysis and statistics. G.F., M.M., and G.S. supervised experiments and critically revised the
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26	

27 Abstract

High-yielding dairy cows are metabolically challenged during transition, when intense mobilization 28 29 and hepatic oxidation of lipids is achieved, thus leading to fatty infiltration, ketosis and generalized 30 inflammation. The condition is associated to periparturient diseases and poor fertility. The aim of 31 this study was to assess whether serum lipoprotein concentrations in the dry period could influence 32 the occurrence of postpartum diseases and reproductive performance in dairy cows. The study was carried out on 30 multiparous Holstein Friesian cows. Blood samples were collected at dry-off (-60 33 34 days), 30 days after dry-off and within 12 hours after parturition for biochemical and serum lipoprotein assays. From 10 to 60 days after parturition milk was collected twice weekly after 35 feeding, for milk whey progesterone assay. The Optimal Cutpoint package identified a threshold of 36 37 89% for serum High Density Lipoprotein (HDL) concentration at the beginning of the dry period 38 with 95% of confidence interval. Cows with serum HDL greater than 89% (High group, n = 10) 39 showed better reproductive performance when compared to those with low values (Low group, n =18). The odds ratio for reproductive disorders in High group was 0.6875, however, differences were 40 41 not significant probably due to both the reduced number of animals per group and overall low 42 incidence of postpartum reproductive disease. First postpartum luteal activity occurred around day 43 23, while the second one between days 40 and 48. The average calving to first AI interval was 64.00 \pm 3.95 days and 94.50 \pm 12.32 days in High and Low group, respectively (P < 0.05). The calving-44 45 conception interval was 129.86 ± 24.42 days and 199.18 ± 24.73 days in High and Low groups, respectively (P < 0.05). Low group displayed an increase in liver markers, that is total bilirubin, 46 47 with $0.46 \pm 0.09 \text{ mg/dL}$ and $0.23 \pm 0.08 \text{ mg/dL}$, in Low and High group respectively (P < 0.05), and NEFA/cholesterol ratio, with 0.30 ± 0.06 and 0.14 ± 0.03 , in Low and High groups, respectively (P 48 49 < 0.05), at parturition. Concentrations of HDL > 89% at dry-off could be suggestive of improved 50 liver adaptation to the transition, and probably of enhanced fertility in High group.

51

52 Key words: dairy cow; dry period; lipoprotein; fertility; reproductive disorders

53 **1 Introduction**

In the last weeks of gestation, cow's metabolism has to sustain fetal growth, mammary gland 54 development and after calving the energy demand further increases due to lactation. Usually, during 55 this period the dry matter intake is not sufficient to meet the requirements. In order to cope with this 56 57 imbalance, dairy cows mobilize great amounts of body reserves, especially lipids [1]. Lipids are 58 released from periphery into the circulation in the form of non-esterified fatty acids (NEFA) and 59 then they are used in liver for gluconeogenesis and ketogenesis. If negative energy balance is 60 excessive, the intake of NEFA overcomes the possibility of complete oxidation in the liver. In this 61 case, NEFA are re-esterified, turned into triglycerides and stored into the cytoplasm of hepatocytes as lipid droplets. This condition, known as fatty liver, is usually associated to impairment of liver 62 63 function, subclinical or clinical ketosis, periparturient metabolic diseases and poor fertility, which deeply influence the herd profitability [2-5]. Human and bovine hepatocytes are able to synthesize 64 65 very low-density lipoprotein (VLDL), in order to export triglycerides towards peripheral tissues [6]. However, bovine liver is not able to adjust VLDL synthesis based on NEFA absorption and re-66 67 esterification into cytoplasmic droplets [2]; therefore, the prevalence of lipidosis among transition 68 dairy cows can reach 50% [7-8]. Very-low density lipoprotein represents only 3 to 5% of all 69 circulating lipoproteins in cattle, while other classes, such as high-density (HDL) and low-density 70 lipoprotein (LDL), account for 80-85% and 10-15%, respectively. As liver can export triglycerides 71 only through VLDL, it is generally assumed that low levels of circulating VLDL are representative 72 of increased fatty acids infiltration [6, 9].

It has been reported a decrease of pregnancy at first artificial insemination when more than 50% of cows in a herd have serum NEFA ≥ 0.5 mEq/L one week before calving, as indicative of intense lipids mobilization [3]. An increased odds ratio for retention of fetal membranes and metritis in cows with prepartum NEFA ≥ 0.3 mEq/L was reported [4]. It has also been highlighted that the peculiar metabolic status of the dairy cows during periods of lipids mobilization could cause lipid accumulation in oocytes and the regenerating endometrium, which impairs fertility via reduction in 3

embryo survival and increased inflammatory changes, respectively [10]. The majority of studies
concerning the association between biochemistry profiles of dry cows in late gestation and
postpartum performance are focused on the last two or three weeks before calving [3-4, 9].
However, Dervishi et al. [11] reported alterations in inflammatory and metabolic profiles as early as
eight weeks before calving, in dairy cows that will develop metritis. Brscic et al. [12], recently
reported some reference limits for metabolic profiles in Holstein late-pregnant heifers and dry cows,
but postpartum performance was not assessed.

Moved by the need of finding new predictive indexes of reduced reproductive efficiency, we hypothesized that the level of circulating lipoproteins during the last period of pregnancy could be indicative of the degree of adaptation to transition of dairy cows and of their susceptibility to both postpartum reproductive diseases and reduced fertility. Moreover, we evaluated biochemistry profiles from dry-off until calving as a tool to verify the overall health status in our experimental animals and to exclude alterations in lipoprotein metabolism due to subclinical pathological conditions.

93

94 **2 Materials and Methods**

95 2.1 Animals and husbandry

In the present study, 30 healthy multiparous Holstein Friesian cows were selected with parity 96 97 ranging from 2 to 5; they belonged to a commercial dairy farm located in Central Italy (42°95' N, 12°39' E), with a herd size ranging between 100 and 120 lactating cows. Each cow was randomly 98 99 selected through the herd-management software before entering into the dry period. Mean heard 100 intercalving period was 410.27 ± 15.87 days, the mean dry period length was 60.25 ± 10.14 days 101 and the voluntary waiting period averaged 50.16 ± 7.08 days. Average milk production was above 102 10,900 kg/lactation/cow. All lactating cows were housed in free stalls with cubicles and milked with 103 two Automatic milking systems (DeLaval S.p.A., VMS, Milano, Italy); dry cows were kept in a free 104 stall barn with straw. The dry cows had free access to a total mixed ration (TMR) offered ad

105 *libitum*, composed of 4.5 kg wheat straw (4.60% CP, 78.90% NDF, 48.40% ADF), 4.5 kg oat hay 106 (8.70% CP, 61.30% NDF, 38.20% ADF), 3.0 kg concentrate (28.50% CP, 20.20% NDF, 9.70% 107 ADF) per head. During the close-up period this ration was supplemented with 10 kg of fresh cow 108 TMR/head. The overall TMR composition was: 4 kg alfalfa and lolium mixed hay (7.67% CP, 109 37.86% NDF, 20.84% ADF), 3.5 kg alfalfa hay (14.91% CP, 42.70% NDF, 33.10% ADF), 11.5 kg concentrate (28.50% CP, 20.20% NDF, 9.70% ADF), 22 kg corn silage (9.20% CP, 45.90% NDF, 110 111 27.00% ADF) and 0.8 kg molasses (4.30 % CP, 0% NDF, 0% ADF). Mycotoxins content in feed 112 was within the legislative established limits (Italian Law 149/2004). Contents of organic acid in 113 silage and total mixed ration from dry cow feed-bunk were evaluated by HPLC analysis. Only 114 butyric acid was fairly present in both silage and total mixed ration, with mean values of $0.02 \pm$ 115 0.01 and 0.03 ± 0.001 g/100g, respectively.

116

117 2.2 Experimental procedures and samples collection

118 The study was conducted from January to December 2015; all experimental cows calved before the 119 end of April 2015. Two cows were excluded from the experiment due to abortion and premature 120 calving. Body condition score (BCS), assessed through a five-point scale [13], and blood sampling 121 were performed at 60 (T1) and 30 days (T2) before the expected calving and within 12 hours 122 postpartum (T3). Samples were obtained by coccygeal vein puncture into plain and EDTA vacuum 123 tubes (BD Vacutainer Systems, Plymouth, UK) between 5.30 and 6.00 AM, half an hour before 124 feeding. Samples were stored at 4°C and delivered within 1 hour to the Laboratory; serum was 125 obtained through centrifugation at 1,300 g for 10 minutes and immediately processed. An aliquot of serum from each sample was stored at 4°C until electrophoresis analysis. From 10 to 60 days after 126 127 parturition milk was collected twice weekly after feeding, between 6.00 and 7.00 AM, in empty eppendorf tubes and immediately frozen at -20°C, until analysis. 128

129 To evaluate healthy condition of the newborn calf, the APGAR score described by Vannucchi et al.

- 130 [14] was employed. Briefly, the following symptoms were considered and scored: mucous color (0
- 131 = cyanotic; 1 = pale; 2 = normal), heart rate (0 = absent; 1 = bradycardia, < 80 beats per minute or
- 132 irregular; 2 = regular, > 100 beats per minute), muscle tone (0 = flaccid; 1 = slight flexion; 2 =
- 133 flexion), activity (0 = absent; 1 = some movement; 2 = active calf), and respiration (0 = absent; 1 =
- 134 irregular < 24 respiration per minute; 2 = regular > 36 respiration per minute).
- 135 Postpartum uterine diseases were diagnosed according to Sheldon et al. [15]. Briefly, animals were
- 136 monitored once weekly after calving until complete uterine involution was achieved. The fetal
- 137 membranes were considered retained when not released within 12 hours after parturition. Cows that
- 138 presented an enlarged uterus with watery red-brown to viscous off-white purulent uterine discharge,
- 139 from 0 to 21 days after calving were considered suffering from metritis. Clinical endometritis was
- 140 defined by the presence of pathological uterine discharge 21 days or more postpartum.
- 141 Reproductive parameters such as calving to first insemination interval, calving to conception
- 142 interval and number of inseminations per pregnancy were retrieved from the herd management
- 143 software, at least until 200 days after parturition.
- 144 The experimental activity was carried out in accordance to the guidelines of animals experiments as
- set by the Italian Law 26/2014 (national application of EU Directive 2010/63/EU) and has been

146 approved by the Ethical Committee of the University of Perugia.

147

148 2.3 Serum lipoprotein's electrophoresis, biochemical profiles and CBC

149 Electrophoresis of serum lipoproteins was obtained with the Hydrasys – LC Sebia automatic system

150 and Hydragel 7 LIPO + Lp(a) Kit (Sebia® Electrophoresis, Sebia Inc., Norcross, GA USA). Briefly,

- 151 lipoprotein classes were separated by electrophoresis on agarose gel buffered plates (pH 8.5);
- different bands were then read with densitometry scanner at 570 nm (Epson Perfection V700
- 153 PHOTO, Seiko Epson Corporation, Japan) and lipoproteins were expressed in term of relative
- 154 percentage. Three main bands were identified, namely HDL (or α-lipoproteins), VLDL (or pre-β-
- 155 lipoproteins) and LDL (or β -lipoproteins).

- 156 The serum concentrations of albumin (ALB), β -hydroxybutyrate (BHB), total proteins (TP),
- 157 triglycerides (TG), total bilirubin, direct bilirubin, indirect bilirubin, glucose, urea, creatinine, lactic
- 158 dehydrogenase (LDH), aspartate transaminase (AST), alanine transaminase (ALT), non-esterified
- 159 fatty acids (NEFA), gamma-glutamyl transpeptidase (GGT), alkaline phosphatase (ALP), creatine
- 160 kinase (CK), cholesterol (CHOL), magnesium (Mg), calcium (Ca) and phosphorus (P) were
- 161 determined with an Hitachi 904 chemistry analyzer (Boehringer Mannheim, Germany).
- 162 All diagnostic kits were purchased from DiaSys Diagnostic Systems (GmbH, Alte Strasse 9, D –
- 163 65558 Holzheim, Germany). Count of blood cells (CBC) was obtained in EDTA samples with a
- 164 Heco Vet S (SEAC, Florence, Italy) electric impedance system.
- 165

166 2.4 Milk whey progesterone assay

- Milk progesterone was quantified into the whey fraction through direct enzyme immunoassay as
 described by Comin et al. [16]. Briefly, whey aliquots were added to micro plates, with 25 µL/well
- 169 of anti-progesterone antibody with 50 µL of progesterone-peroxidase conjugate in ELISA buffer.
- 170 Plates were incubated one night at 4°C and then washed five times with buffer solution.
- 171 Chromogenic substrate was added and plates were incubated for 30 minutes at room temperature.
- 172 Finally, absorbance was read at 450 nm using a plate reader. Resumption of ovarian activity was
- 173 considered when progesterone concentrations was above 300 pg/mL [16], and presence of a corpus
- 174 luteum was confirmed by trans rectal ultrasound examination.
- 175

176 2.5 Statistical analysis

177 Data were analyzed through SPSS software [17]. Normal distribution was assessed for each

- 178 biochemical and reproductive outcome through Kolmogorov-Smirnov test with 95% of confidence
- 179 interval. However, due to the limited number of animals involved and in order to be conservative,
- 180 non-parametric statistics were used. All tests were set with $\alpha < 0.05$. Data were expressed through
- 181 mean and standard error of the mean (SEM). Differences between groups were analyzed with

182 Kruskal-Wallis test. R software v3.2.4 [18] was used to run Optimal Cutpoints package [19] in order to estimate threshold in lipoproteins concentration at T1, T2 and T3 in relation to postpartum 183 184 diseases. The optimal cutpoint for the diagnostic test (HDL, VLDL and LDL in each of the three periods) was computed by means of a Receiver Operating Characteristic (ROC), using the disease 185 186 status (e.g. sick or healthy) as dependent categorical variable and optimizing the Younden's Index, 187 so that the sum of Specificity and Sensitivity was maximized. Cows were then assigned to Low and High HDL groups; differences in reproductive outcomes and mean progesterone concentrations 188 189 were investigated with Kruskal Wallis test in SPSS. If any significant difference was found between 190 High and Low HDL cows, data were discussed as belonging to a single group based on sampling 191 interval.

192

3 Results

194 *3.1 Clinical outcomes and resumption of ovarian activity*

195 Cows entered the dry period with a mean BCS of 3.23 ± 0.07 that decreased to 3.04 ± 0.05 at

196 calving (P < 0.05). Nineteen out of 28 animals had normal postpartum while 3 cows showed

197 retained fetal membranes and 6 metritis. All calving were eutocic and all calves were born healthy

and with a mean APGAR score of 8.21 ± 1.15 .

199 Left abomasal displacement was observed in 3 animals without any reproductive disorder.

200 Milk whey progesterone in High (n = 10) and Low (n = 18) group of cows ranged between 150 and

201 500 pg/mL, but no significant difference was identified. First postpartum luteal activity occurred

around day 23, while the second one from day 40 to 48. One animal in High and one in the Low

203 group revealed an increase in P4 concentrations only after 50 days post calving. One cow per group

also showed a prolonged period of elevated milk P4.

- 205
- 206

207 3.2 Biochemistry profiles and CBC

208 3.2.1 Serum lipoprotein electrophoresis

209 Relative proportion of lipoproteins in experimental dairy cows is reported in Table 1. VLDL

- lipoproteins ranged between 3.27 ± 0.29 % and 2.69 ± 1.43 %, without any difference between
- 211 groups; HDL decreased (P < 0.001) from dry-off (87.52 ± 0.87 %) towards T2 (82.41 ± 1.21 %),
- then increased at calving $(87.39 \pm 1.07 \%)$. LDL lipoproteins presented a reverse trend; their
- 213 concentration was greater at T2, with mean value of 14.32 ± 1.18 % (P < 0.01).
- 214 The Optimal Cutpoint package identified a threshold of 89% for serum HDL concentration at the
- beginning of the dry period with 95% of confidence interval. As shown in Table 2, cows with serum
- HDL greater than 89% (n = 10) tended to show better reproductive performances when compared to
- others (n = 18). More deeply, the average calving to first AI interval was 64.00 ± 3.95 days and
- 94.50 ± 12.32 days in High and Low group, respectively (P < 0.05). The calving conception
- intervals were 129.86 ± 24.42 days and 199.18 ± 24.73 days in High and Low groups, respectively
- 220 (P < 0.05). The odds ratio for reproductive disorders in High HDL cows was 0.6875, even if the P
- 221 was not significant.
- 222

223 *3.2.2 Energy metabolism analytes*

Beta-hydroxybutyrate, glucose, NEFA and triglyceride concentration are reported in Table 3. Betahydroxybutyrate was constantly below reference limit of 1.40 mmol/L and did not show any significant variation throughout the period of study. Mean serum glucose level decreased from $66.11 \pm 1.85 \text{ mg/dL}$ at T1 to $55.75 \pm 2.24 \text{ mg/dL}$ at parturition (P < 0.001) while NEFA increased from $0.18 \pm 0.02 \text{ mmol/L}$ at dry-off to $0.58 \pm 0.08 \text{ mmol/L}$ at calving (P < 0.001). Triglycerides increased both at T1 and T2, while they declined at calving time (P < 0.001).

230

231 3.2.3 Markers of liver function

232 Low group displayed significantly higher concentration of both total and indirect bilirubin, and

233 NEFA/CHOL ratio than High group, only at calving (Table 4).

234 In all cows, urea and LDH did not show any significant variation during the dry period. Aspartate 235 transaminase activity was higher at calving, with a mean value of 106.57 ± 6.98 UI/L, compared to 236 what found during the dry period (P < 0.001). Concerning GGT, it decreased at parturition with a mean value of 15.22 ± 1.31 UI/L (P < 0.05) and ALT showed a similar trend, averaging $20.21 \pm$ 237 238 0.96 UI/L at calving (P < 0.001). Cholesterol declined progressively from 192.79 ± 5.30 mg/dL to 239 $112.57 \pm 6.96 \text{ mg/dL}$ (P < 0.001; Table 5). 240 241 3.2.4 Protein metabolism analytes 242 Protein metabolism analyses are reported in Table 6. Albumin and CK did not show any significant

variation throughout the period of study. Conversely, total serum proteins decreased from T1 (7.56 ± 0.16 g/dL) until parturition, when they reached an average value of 6.43 ± 0.11 g/dL (P < 0.001); CK remained constant and averaged 265.69 \pm 52.59 UI/L throughout the study.

246

247 *3.2.5 Mineral metabolism and other analytes*

Mineral and other analytes are reported in Table 7. At dry-off, calcium and phosphorus averaged 9.65 \pm 0.12 mg/dL and 6.55 \pm 0.28 mg/dL, respectively; thereafter they decreased at calving, with mean values of 8.80 \pm 0.29 mg/dL (P < 0.05) and 4.83 \pm 0.28 mg/dL (P < 0.001), respectively. Alkaline phosphatase activity was higher at dry-off, with a mean value of 84.57 \pm 4.68 UI/L and then decreased to 64.46 \pm 5.04 UI/L at calving (P < 0.001). Creatinine constantly averaged 1.25 \pm 0.06mg/dL. Magnesium and Creatinine did not show any significant variation throughout the period

of study.

255

256 *3.2.6 Count blood cells*

The CBC features were within the referenced limits [20] and there were any significant differenceamong groups (data not shown).

259

261 4 Discussion

262 The transition dairy cow has to cope with negative energy balance, lipomobilization and fatty liver,

all of them influencing the susceptibility to postpartum diseases and the reproductive outcomes.

However, there is lack of data concerning lipoprotein metabolism during the full dry period and

their interaction with postpartum health and fertility.

High-density lipoproteins, herein reported, were greater at dry-off and at calving, compared to T2, 266 267 while LDL reached a peak one month before expected delivery. This opposite trend could be due to 268 LDL peripheral catabolism, as mammary gland absorbs triglycerides from LDL for milk fatty acids 269 synthesis; in the middle of dry period, this metabolism is not active and LDL is accumulated, thus 270 determining a relative decline of HDL concentration. We also observed a progressive decline of 271 cholesterol until reaching the lower values at calving, as also previously reported [7, 12, 21-24]. 272 Generally, it is accepted that cholesterol is representative of liver synthesis of VLDL and that a 273 decline in VLDL concentration is indicative of poor adaptation of the liver to fat mobilization, even 274 if currently, there is no reference limit for VLDL in dry high-producing dairy cows [9, 21, 25-26]. 275 Through Optimal Cutpoint package, we defined a cut-off value of 89% of serum HDL at the 276 beginning of the dry period as a possible threshold to identify cows, which are more prone to 277 experience postpartum reproductive diseases. Cows belonging to High group presented better 278 calving to first AI and calving to conception intervals when compared to Low group (P<0.05). 279 Cows with a greater concentrations of HDL had also lower incidence of retention of fetal 280 membranes and/or metritis, with an odds ratio of 0.6875; however, differences were not significant 281 probably due to both the reduced number of animal per group and the overall low incidence of 282 postpartum reproductive diseases. In the present study, in fact, the prevalence of fetal membrane 283 retention and/or metritis was 32%, which was lower than the rate of 36-50% reported by Sheldon et al. [15]. At parturition, bilirubin and NEFA/CHOL ratio were higher in Low group, as indicative of 284 285 increased liver metabolic load, even if values remained within reference limits [9, 22]. 11

All cows resumed ovarian activity within the first 3 week s postpartum. Only one cow per group showed a prolonged period of milk P4 due to persistent ovarian luteal cyst. Emergence and growth of first follicular wave after calving is regulated by nutritional status and energy availability which also influence hypothalamic release of gonadotropin releasing hormone [27]. Since resumption of ovarian activity was similar between groups, we hypothesized that serum lipoproteins did not influence the hypothalamic-pituitary-gonadal axis.

292 The reduction in reproductive efficiency observed in the Low group could be related to oocyte-293 embryo quality. Considering that dairy cows have to cope with generalized inflammatory condition 294 4-5 weeks before calving [23] and that HDL have anti-inflammatory activity [29], we hypothesized that their high concentration in dry cows could protect primordial ovarian follicles against 295 296 metabolic stress, enhancing oocyte quality, and increasing fertility rate, as observed in High group. 297 In relation to postpartum follicular development and their steroidogenic capacity, Golini et al. [28] 298 showed that there is a down-regulation of genes involved in cell proliferation and steroidogenesis of granulosa cells harvested from preovulatory follicles collected from early lactating cows, until 90 299 300 days after calving. Moreover, results reported by Wu et al. [30] show that ovarian cells 301 preferentially use HDL-bound cholesterol for steroidogenesis, suggesting their fundamental role in 302 follicle development and maturation. Even if those results are referred to active and growing 303 follicles, it could be speculated that decreased HDL availability in conjunction with lower follicle's 304 steroidogenic capacity and pro-oxidant environment could negatively affect cholesterol absorption 305 storage and metabolism in ovarian follicles during the dry period.

To our knowledge, only other two studies reported data about concentration of lipoproteins in dairy
cows during dry period [7, 29]. The lack of uniformity in separation and detection methods

308 (electrophoresis *versus* centrifugation) or the investigation of lipoprotein concentration during

309 lactation makes comparison with some studies difficult [25, 31]. Bernabucci et al. [7] used the same

analytical procedure as in the present study, but they considered a different period of samples

311 collection (35 days before calving and 3 and 30 days after calving). They reported a lower

312 proportion of serum VLDL and higher HDL after calving, while LDL did not significantly change 313 during the transition period. This difference could be due to the matrix used for the test, since 314 Bernabucci et al. [7] worked on plasma while Sebia Lipo Lp (a) electrophoresis method is 315 standardized for serum. Newman et al. [29] investigated the effect of several diets with different 316 energy density on serum lipoprotein composition, from week 6 before expected calving until week 317 6 of lactation. The study [29] was aimed to verify if a provision of adequate total metabolizable energy would better allow the cow to maintain total cholesterol and an HDL-rich profile throughout 318 319 the transition period while concurrently limiting the rise in NEFA. However, they did not find 320 influence of energy density in the diet on lipoprotein. This suggests that lipoprotein metabolism is dependent on liver and peripheral secretion and absorption rather than on feeding management. 321 322 They reported a decline in HDL from week 6 until week 1 before calving and a subsequent increase. 323 In their study, mean concentrations of HDL at the beginning and at the end of the dry period were 324 similar to those found in the present work, for instance 80-85%. Since in their study VLDL was not 325 detectable, LDL relative concentrations were complementary to that of HDL, thus ranging from 326 15% to 25%. This difference could be due to the methods used, such as the duration of 327 electrophoresis and migration, the voltage applied or the staining technique, which may affect the 328 degree of lipoprotein separation and identification.

329 All other analysis carried out to verify animal's overall health status and to exclude alterations in 330 lipoprotein metabolism due to subclinical pathological conditions confirmed the good health 331 condition of the cows. Only during the last month before calving some modifications, indicated a 332 certain degree of lipid mobilization and reactivation of liver function, typical of the dairy cows 333 during the transition period. In particular, BCS was evaluated to assess the overall energy balance 334 and it was fairly constant, while mean NEFA were within reference limit of 0.4 mmol/L, as reported 335 in pending calving cows [4, 9, 32]. Only at calving BCS significantly decreased, as generally described in transition dairy cows [4, 24, 33-34]. Correspondingly, NEFA increased, but remained 336 337 lower than the threshold value of 0.7 mmol/L established for the postpartum cows [32]. Triglyceride

338	decreased significantly from dry-off to calving, as previously observed by Weber et al. [24]. Herein,
339	cows showed normal glucose concentrations throughout the dry period, similarly to what reported
340	by Weber et al. [24]; however, a significant decrease was found at parturition. We observed BHB
341	was constantly greater than 1.00 mmol/L but below the threshold value of 1.20 - 1.40 mmol/L. Feed
342	analysis excluded the presence of ketogenetic compounds, as only butyric acid was fairly present in
343	both silage and total mixed ration. Subclinical ketosis diagnosis has been defined by BHB
344	concentrations of 1200 or 1400 μ mol/L [35-37]. Although BHB above 0.96 mmol/L is associated
345	with increased risk of postpartum disease [38-39], prepartum BHB concentrations are not predictive
346	of disease [4].
347	Concerning the other analytes, mean concentrations of GGT, ALT and TP were in agreement with
348	results reported by Brscic et al. [12], without significant difference between groups. The increase of
349	mean AST levels toward calving in all our experimental groups could be representative of increased
350	muscle labour. As ALP, Ca and P concentrations decreased at parturition but remained within

351 reference limits; we excluded subclinical hypocalcemia or other mineral imbalance.

352

353 **5 Conclusions**

354 The growing knowledge on the role of lipid metabolism in the adaptation of dairy cows to the 355 transition lead to an increased interest in lipoproteins. In our study, animal with HDL levels greater 356 than 89% at dry-off showed better reproductive performance when compared to the other experimental group. We suggest that HDL due to their role of main carrier of cholesterol in ovarian 357 358 steroidogenetic cells and to anti-inflammatory and antioxidant activities can exert some influences 359 on liver adaptation to the transition and on the fertility in cows. However, future studies are needed to improve our knowledge on lipoprotein metabolism and to validate the quantification of HDL in 360 dry and transition phases using larger groups of animals and under different management or 361 nutritional conditions. 362

364 6 References

- [1] Bobe G, Young JW, Beitz DC. Invited review: pathology, ethiology, prevention, and treatment of
 fatty liver in dairy cows. J Dairy Sci 2004;87:3105–24.
- 367 [2] Bauchart D. Lipid absorption and transport in ruminants. J Dairy Sci 1993;76:3864–81.
- 368 [3] Chapinal N, LeBlanc SJ, Carson ME, Leslie KE, Godden S, Capel M et al. Herd-level
- 369 association of serum metabolites in the transition period with disease, milk production and early
- 370 lactation reproductive performance. J Dairy Sci 2012;95:5676–82.
- 371 [4] Ospina PA, McArt JA, Overton TR, Stokol T, Nydam DV. Using nonesterified fatty acids and β-
- 372 hydroxybutyrate concentrations during the transition period for herd-level monitoring of increased
- 373 risk of disease and decreased reproductive and milking performance. Vet Clin North Am Food Anim
- 374 Pract 2013;29:387–412.
- 375 [5] Sordillo LM, Raphael W. Significance of metabolic stress, lipid mobilization, and inflammation
 376 on transition cow disorders. Vet Clin North Am Food Anim Pract 2013;29:267–78.
- [6] Katoh N. Relevance of apolipoproteins in the development of fatty liver and fatty liver-related
 peripartum diseases in dairy cows. J Vet Med Sci 2002;64: 293–307.
- 379 [7] Bernabucci U, Ronchi B, Basiricò L, Pirazzi D, Rueca F, Lacetera N et al. Abundance of mRNA
- 380 of apolipoprotein b100, apolipoprotein e, and microsomal triglyceride transfer protein in liver from
- 381 periparturient dairy cows. J Dairy Sci 2004;87:2881–8.
- 382 [8] Roche JR, Kay JK, Friggens NC, Loor JJ, Berry DP. Assessing and managing body condition
- score for the prevention of metabolic disease in dairy cows. Vet Clin North Am Food Anim Pract
 2013;29:387–412.
- 385 [9] Van Saun RJ. Metabolic profiling of transition dairy cows: can we predict impending problems?
- 386 Proceedings Danske Kvægfagdyrlægers Arsmode (Danish Bovine Practitioner Seminar),
- 387 Middlefart, Denmark, January 2007;24–5.
- 388 [10] Wathes DC, Clempson AM, Pollot GE. Association between lipid metabolism and fertility in
- the dairy cow. Reproduction, Fertility and Development, 2013;25, 48-61.

- 390 [11] Dervishi E, Zhang G, Hailemariam D, Goldansaz SA, Deng Q, Dunn SM, Ametaj BN.
- 391 Alterations in innate immunity reactants and carbohydrate and lipid metabolism precede occurrence
- of metritis in transition dairy cows. Res Vet Sci. 2016 Feb;104:30-9.
- 393 [12] Brscic M, Cozzi G, Lora I, Stefani AL, Contiero B, Ravarotto L et al. Short communication:
- 394 Reference limits for blood analytes in Holstein late-pregnant heifers and dry cows: Effects of parity,
- days relative to calving, and season. J Dairy Sci 2015;98:7886–92.
- [13] Edmonson AJ, Lean IJ, Weaver LD, Farver T, Webster G. A body condition scoring chart for
 holstein dairy cows. J Dairy Sci 1989;72:68–78.
- 398 [14] Vannucchi CI, Rodrigues JA, Silva LCG, Lúcio CF, Veiga GAI. Effect of dystocia and
- 399 treatment with oxytocin on neonatal calf vitality and acid-base, electrolyte and haematological
- 400 status. The Veterinary Journal 2015;203:228-232.
- 401 [15] Sheldon IM, Cronin J, Goetze L, Donofrio G, Schuberth HJ. Defining postpartum uterine
- 402 disease and the mechanisms of infection and immunity in the female reproductive tract in cattle
- 403 Biol Reprod 2009;81:1025–32.
- 404 [16] Comin A, Renaville B, Marchini E, Maiero S, Cairoli F, Prandi A. Technical note: direct
- 405 enzyme immunoassay of progesterone in bovine milk whey J Dairy Sci 2005;88:4239–42.
- 406 [17] SPSS Incorporated: Statistic Package for Social Science (SPSS) Version 16 Chicago, IL USA:
- 407 SPSS Inc; 2007.
- 408 [18] RStudio, R software v3.2.4, https://www.rstudio.com/ [accessed 24.03.16].
- 409 [19] Lopez-Raton M, Rodriguez-Alvarez MX Package OptimalCutpoints https://cran.r-
- 410 project.org/web/packages/OptimalCutpoints/OptimalCutpoints.pdf. [accessed 24.03.16].
- 411 [20] Smith BP. Large animal internal medicine. 5th ed. Mosby Elsevier. 2015.
- 412 [21] Stengärde L, Tråvén M, Emanuelson U, Holtenius K, Hultgren J, Niskanen R. Metabolic
- 413 profiles in five high-producing Swedish dairy herds with a history of abomasal displacement and
- 414 ketosis. Acta Veterinaria Scandinavica 2008;50:31.
- 415 [22] Kalaitzakis E, Panousis N, Roubies N, Giadinis N, Kaldrymidou E, Georgiadis M et al.
 16

- 416 Clinicopathological evaluation of downer dairy cows with fatty liver. The Canadian Veterinary
- 417 Journal 2010;51:615–22.
- 418 [23] Trevisi E, Amadori M, Cogrossi S, Razzuoli E, Bertoni G. Metabolic stress and inflammatory
- 419 response in high-yielding, periparturient dairy cows. Res Vet Sci 2012;93:695–704.
- 420 [24] Weber C, Hametner C, Tuchscherer A, Losand B, Kanitz E, Otten W et al. Variation in fat
- 421 mobilization during early lactation differently affects feed intake, body condition, and lipid and
- 422 glucose metabolism in high-yielding dairy cows. J Dairy Sci 2013;96:165–80.
- 423 [25] Sevinç M, Basoglu A, Guzelbektas H. Lipid and lipoprotein levels in dairy cows with fatty
- 424 liver Tusk J Vet Anim Sci 2003;27:295–9.
- 425 [26] Bertoni G, Trevisi E. Use of the liver activity index and other metabolic variables in the
- 426 assessment of the metabolic health in dairy herds. Vet Clin North Am Food Anim Pract
- 427 2013;29:413–31.
- 428 [27] Chagas LM, Bass JJ, Blache D, Burke CR, Kay JK, Lindsay DR et al. Invited review: New
- 429 perspectives on the roles of nutrition and metabolic priorities in the subfertility of high-producing
- 430 dairy cows. J Dairy Sci 2007;90:4022–32.
- 431 [28] Golini VE, Stradaioli G, Sirard MA. Transcriptome analysis of bovine granulosa cells of
- 432 preovulatory follicles harvested 30, 60, 90, and 120 days postpartum. Theriogenology 2014;82:580–
 433 91.
- 434 [29] Newman A, Mann S, Nydam DV, Overton TR, Behling-Kelly E. Impact of dietary plane of
- 435 energy during the dry period on lipoprotein parameters in the transition period in dairy cattle. J
- 436 Anim Physiol Anim Nutr (Berl) 2016;100:118–26.
- 437 [30] Wu Q, Sucheta S, Azhar S, Menon KMJ. Lipoprotein enhancement of ovarian theca-interstitial
- 438 cell steroidogenesis: relative contribution of scavenger receptor class B (Type I) and Adenosine 5'-
- 439 Triphosphate-Binding Cassette (Type A1) transporter in High-density lipoprotein-cholesterol
- 440 transport and androgen synthesis. Endocrinology 2003;144(6):2437-2446.
- 441

- 442 [31] Ileri-Büyükoğlu T, Sahinduran S, Sezer K, Güldür T. Evaluation of changes in serum
- 443 lipoprotein and apolipoprotein patterns in cows with ketosis. Am J Vet Res 2009;70:563–70.
- 444 [32] Mulligan FJ, O'Grady L, Rice DA, Doherty ML. A herd health approach to dairy cow nutrition
- and production diseases of the transition cow. Animal Reproduction Science 2006;96:331–53.
- 446 [33] Ingvartsen, KL and Andersen JB. Integration of metabolism and intake regulation: A review
- 447 focusing on periparturient animals. J Dairy Sci 2000;83:1573–97.
- 448 [34] Drackley JK, Overton TR and Douglas GN. Adaptations of glucose and long-chain fatty acid
- 449 metabolism in liver of dairy cows during the periparturient period. J Dairy Sci 2001;84:E100–E112.
- 450 [35] Duffield TF, Lissemore KD, McBride BW, Leslie KE. Impact of hyperketonemia in early
- 451 lactation dairy cows on health and production. J Dairy Sci 2009;92:571–80.
- [36] LeBlanc S. Monitoring metabolic health of dairy cattle in the transition period. J Reprod Dev
 2010;56:S29–S35.
- 454 [37] Chapinal N, Carson ME, LeBlanc SJ, Leslie KE, Godden S, Capel M et al. The association of
- 455 serum metabolites in the transition period with milk production and early-lactation reproductive
- 456 performance. J Dairy Sci 2012;95:1301–09.
- 457 [38] Roussel JD, Seybt SH, Toups G. Metabolic profile testing for Jersey cows in Louisiana:
- 458 reference values. Am J Vet Res 1982;43:1075–7.
- 459 [39] Eicher R. Metabolic profile testing in dairy herds: wrong answer or wrong question? Acta
- 460 Veterinaria Scandinavica 2003;44(Suppl 1):28.
- 461

%	Time	Mean ± SEM	Р
	T1	87.52 ± 0.88^{a}	
High-density lipoproteins	T2	82.41 ± 1.21^{b}	P < 0.001
	Т3	87.39 ± 1.07^{a}	
	T1	$9.65\pm0.71^{\text{a}}$	
Low-density lipoproteins	T2	14.32 ± 1.18^{b}	P < 0.01
	Т3	9.92 ± 1.10^{a}	
	T1	2.83 ± 0.29	
Very low-density lipoproteins	T2	3.27 ± 0.29	NS
	Т3	2.69 ± 1.43	

462 Table 1. Relative proportion of lipoproteins in the 28 experimental dairy cows.

T1 = 60 days before expected calving; T2 = 30 days before expected calving; T3 = calving; P =

464 Kruskal-Wallis test significance; NS = not significant. ^{a,b} = Values with different superscripts are
465 significantly different.

	Cut-off HDL > 89.0 %						
		n	HDL (%)	C - 1AI (days)	C-C (days)	AI (n°)	OR
	Low	18	85.07 ± 0.88	94.50 ± 12.32	199.18 ± 24.73	3.36 ± 0.58	
	High	10	91.94 ± 0.67	64.00 ± 3.95	129.86 ± 24.42	3.67 ± 0.78	0.6875
	Р		P < 0.001	P < 0.05	P < 0.05	NS	NS
471	n = numt	per of anim	nals; $C - 1AI = cal$	ving to first artific	cial insemination in	nterval; C – C =	= calving to
472	conceptio	on interva	l; n° AI = number	of artificial insen	nination per pregna	ancy; $OR = ode$	ds ratio for
473	postpartu	m uterine	diseases; P = Krus	kal-Wallis test sig	nificance; NS = no	t significant.	
474							

470 Table 2. Mean (± SEM) reproductive outcomes in dairy cows of the Low and High HDL groups.

Table 3. Mean (± SEM) values of energy metabolism analytes in the 28 experimental dairy cows 60
(T1) and 30 (T2) days before expected calving, and at calving (T3).

	Time	Values	Р
	T1	1.16 ± 0.08	
BHB (mmol/L)	T2	1.29 ± 0.85	NS
	Т3	1.03 ± 0.53	
	T1	$0.18\pm0.02^{\texttt{a}}$	
NEFA (mmol/L)	T2	$0.24\pm0.04^{\texttt{a}}$	P < 0.001
	Т3	0.58 ± 0.08^{b}	
	T1	$66.10 \pm 1.85^{\text{a}}$	
Glucose (mg/dL)	T2	62.21 ± 1.73^{a}	P < 0.001
	Т3	55.75 ± 2.24^{b}	
	T1	16.32 ± 1.51^{a}	
Triglycerides (mg/dL)	T2	18.39 ± 1.13^{a}	P < 0.001
	Т3	11.41 ± 1.20^{b}	

 $\overline{P} = Kruskal-Wallis test significance; NS = not significant. a,b = Values with different superscripts are$

⁴⁸¹ significantly different.

486 Table 4. Mean (±SEM) bilirubin and NEFA/CHOL ratio in the experimental dairy cows of Low and

487 High HDL groups at calving.

	n	T-Bil	Ind-Bil	NEFA/CHOL	
Low	18	0.46 ± 0.09	0.27 ± 0.07	0.30 ± 0.06	
High	10	0.23 ± 0.08	0.12 ± 0.05	0.14 ± 0.03	
Р		P < 0.05	P < 0.05	P < 0.05	
n = number of animals; T-Bil = total bilirubin; Ind-Bil = indirect bilirubin; NEFA/CHOL = NEFA:					

	Time	Values	Р
	T1	29.39 ± 1.54	
Urea (mg/dL)	T2	30.54 ± 1.89	NS
	Т3	30.89 ± 4.03	
	T1	21.00 ± 2.48^{a}	
GGT (U/L)	T2	16.71 ± 1.68^{b}	P < 0.05
	Т3	15.22 ± 1.31^{b}	
	T1	883.21 ± 87.59	
LDH (U/L)	T2	1113.04 ± 36.80	NS
	Т3	1174.71 ± 60.14	
	T1	82.36 ± 3.95^a	
AST (U/L)	T2	77.00 ± 3.93^{a}	P < 0.001
	Т3	106.57 ± 6.98^{b}	
	T1	29.21 ± 0.97^{a}	
ALT (U/L)	T2	24.54 ± 1.11^{b}	P < 0.001
	Т3	20.21 ± 0.96^{b}	
	T1	192.79 ± 5.30^{a}	
Cholesterol (mg/dL)	Τ2	148.54 ± 5.72^{b}	P < 0.001
	Т3	$112.57 \pm 6.97^{\circ}$	

Table 5. Mean (\pm SEM) hepatic function analytes values in the 28 experimental dairy cows 60 (T1)

494 and 30 (T2) days before expected calving, and at calving (T3).

495 $\overline{P} = Kruskal-Wallis test significance; NS = not significant. a,b,c = Values with different superscripts$ 496 are significantly different.

497

Table 6. Mean (± SEM) protein metabolism analytes values in the 28 experimental dairy cows 60

	Time	Values	Р
	T1	4.10 ± 0.08	
Albumin (g/L)	T2	4.04 ± 0.08	NS
	Т3	4.05 ± 0.08	
	T1	7.56 ± 0.16^a	
Total Proteins (g/L)	T2	7.11 ± 0.13^{a}	P < 0.001
	Т3	6.43 ± 0.11^{b}	
	T1	246.96 ± 59.40	
CK (U/L)	T2	340.34 ± 144.31	NS
	Т3	209.76 ± 28.52	

499 (T1) and 30 (T2) days before expected calving, and at calving (T3).

500 $\overline{P = Kruskal-Wallis test significance; NS = not significant.^{a,b} = Values with different superscripts are$

501 significantly different.

502

	Time	Values	Р
	T1	1.16 ± 0.03	
Creatinine (mg/dL)	T2	1.22 ± 0.05	NS
	Т3	1.38 ± 0.18	
	T1	84.57 ± 4.68^{a}	
ALP (U/L)	T2	$76.07\pm5.72^{\rm a}$	P < 0.001
	T3	64.46 ± 5.05^b	
	T1	$9.65\pm0.12^{\rm a}$	
Calcium (mg/dL)	T2	$9.34\pm0.21^{\text{a}}$	P < 0.05
	T3	8.80 ± 0.29^{b}	
	T1	2.51 ± 0.11	
Magnesium (mg/dL)	T2	2.31 ± 0.08	NS
	Т3	2.39 ± 0.62	
	T1	6.55 ± 0.28^{a}	
Phosphorous (mg/dL)	T2	$6.40\pm0.17^{\text{a}}$	P < 0.001
	Т3	$4.83\pm0.28^{\text{b}}$	

Table 7. Mean (± SEM) mineral metabolism and non-specific analytes values in the 28 experimental
dairy cows 60 (T1) and 30 (T2) days before expected calving, and at calving (T3).

 $\overline{P = Kruskal-Wallis test significance; NS = not significant.^{a,b} = Values with different superscripts are$ 508 significantly different.