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Antioxidant properties of different products and additives in white wine

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Abstract: Different winemaking products (ascorbic acid, glutathione, yeast lees and a yeast autolysate) were tested in comparison with sulfur dioxide, concerning radical scavenging activity (measured by DPPH* assay), oxygen consumption capacity and ability to reduce wine color and predisposition to browning. Trials were performed in white wines and model solution. SO2 was the most active in reducing wine color development. Fresh lees and ascorbic acid were very effective in oxygen and free radical scavenging, but they both induced browning during wine storage, the former, by releasing phenolic compounds. Glutathione was also able to scavenge DPPH* in wine, but less effective against oxygen, and it induced browning during storage. Surprisingly, the yeast derivative preparation was the treatment that behave more similarly to sulfiting; it was very active in scavenging DPPH*, and, even without modifying oxygen consumption rate, it protected quite well wine color and phenolics over a eight months storage time.

Cover Letter

Sulfur dioxide is considered one of the most important enological additives, but it is also toxic and allergenic. In Europe, it is mandatory to label wines with the indication "contains sulfites", if total sulfur dioxide is higher than 10 mg/L; this has determined an increasing interest of the consumers towards the health-related aspects connected with wine consumption. For this reason, the reduction of the use of sulfur dioxide is considered one of the key objectives of modern winemaking and it is strictly connected with the priorities established by the "International Organization of Vine and Wine" (O.I.V.). Current knowledge and technologies allow to postpone sulfiting at the end of alcoholic and malolactic fermentation, with a significant reduction of the overall sulfite levels without jeopardizing wine quality. Contrary, it is more difficult to replace SO₂ after the fermentations, because none of the products currently available for complementing the activity of this additive during wine storage and ageing, is as effective as sulfites themselves. Some of these complementary substances, such as ascorbic acid, have been extensively studied, while some other (e.g. glutathione) were less investigated; anyway, maybe paradoxically, the most of the studies available have been published on technical journals, and the most of the few scientific publications available are focused just on one or two alternatives, sometimes without a direct comparison with sulfur dioxide; moreover, the trials have been generally performed in model solution and only few evidences have been collected on wines. This paper aims to investigate the radical scavenging activity and the oxygen consumption capacity of different enological products and additives (ascorbic acid, glutathione, yeast lees and a yeast derivative preparation), in comparison with SO₂, also considering their effect on wine color and predisposition to browning. Trials were performed in model solution and in different wine typologies. We consider these results just as a preliminary investigation, but, as far as we know, this is one of the most comprehensive, direct comparisons, between sulfur dioxide and other additives, reporting effects on real wines. Moreover, surprisingly, the yeast derivative preparation used, was the additive that behave more similarly to sulfur dioxide, demonstrating its ability in protecting wine color over a 8 months storage time. As far as we know, this is the first time this kind of products has been directly compared with sulfites concerning their antioxidant characteristics.

1 Antioxidant properties of different enological products

2 and additives in comparison with sulfur dioxide

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14 **Abbreviated running title:**

15 Antioxidant capacity of enological products compared with sulfites

Abstract

16

17 Different winemaking products (ascorbic acid, glutathione, yeast lees and a yeast autolysate) 18 were tested in comparison with sulfur dioxide, concerning radical scavenging activity (measured by DPPH• assay), oxygen consumption capacity and ability to reduce wine color 19 20 and predisposition to browning. Trials were performed in white wines and model solution. 21 SO₂ was the most active in reducing wine color development. Fresh lees and ascorbic acid 22 were very effective in oxygen and free radical scavenging, but they both induced browning 23 during wine storage, the former, by releasing phenolic compounds. Glutathione was also able to scavenge DPPH• in wine, but less effective against oxygen, and it induced browning during 24 25 storage. Surprisingly, the yeast derivative preparation was the treatment that behave more 26 similarly to sulfiting; it was very active in scavenging DPPH•, and, even without modifying oxygen consumption rate, it protected quite well wine color and phenolics over a eight 27 28 months storage time.

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- KEYWORDS: sulfur dioxide; yeast derivatives; ascorbic acid; glutathione; yeast lees;
- 31 wine

1 Introduction

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34 Despite the mechanisms involved in wine oxidation have been extensively reviewed 35 (Singleton, 1987; du Toit, Marais, Pretorius, & du Toit, 2006; Waterhouse & Laurie, 2006; 36 Oliveira, Ferreira, De Freitas, & Silva, 2011), the protection of wine against oxidative 37 spoilage remains one of the main goals of modern winemaking, becoming particularly critical 38 when low levels of sulfur dioxide are used. The chemistry of this additive in wine has been 39 recently re-written by Danilewicz (2007; 2011) and Danilewicz, Seccombe, & Whelan 40 (2008): they clearly demonstrated that SO₂ does not react directly with oxygen, as previously thought (Ribéreau-Gayon, Dubourdieu, Doneche, & Lonvaud, 2006), but, in presence of 41 42 metal ions, it is able to scavenge hydrogen peroxide and the quinones formed from the 43 oxidation of polyphenols (Danilewicz et al., 2008). Due to the toxicity and allergenic potential 44 of sulfites, different compounds have been proposed for reducing their final concentration in 45 wine, even if, none of them is likewise effective in protecting wine against oxidations. 46 Ascorbic acid (ASC) is the most known among these products; it is able to scavenge hydroxyl 47 radicals (Bradshaw, Barril, Clark, Prenzler, & Scollary, 2011) and quinones (Waterhouse & Laurie, 2006; Bradshaw et al., 2011), but its metal catalyzed oxidation produces hydrogen 48 49 peroxide (Zoecklein, Fugelsang, Gump, & Nury, 1995; Moreaux, Birlouez-Aragon, & 50 Ducauze, 1996; Ribéreau-Gayon et al., 2006; Bradshaw et al., 2011) and this may trigger 51 browning reactions if sulfites are not present (Bradshaw, Prenzler, & Scollary, 2001; Bradshaw, Cheynier, Scollary, & Prenzler, 2003). The capacity of ascorbic acid to act both as 52 antioxidant and free-radical initiator is known as "crossover effect" (Buettner & Jurkiewicz, 53 54 1996; Bradshaw et al., 2001; Bradshaw et al., 2003) and explains the reason why ASC is normally used in wine in combination with sulfites (Zoecklein et al., 1995; Ribéreau-Gayon et 55 56 al., 2006; Bradshaw, et al., 2011).

Serradilla & Luque de Castro, 2008). Fresh lees have a high oxygen consuming capacity 58 59 (Fornairon-Bonnefond & Salmon, 2003), due to the presence of yeast membrane lipids and 60 sterols (Salmon, Fornairon-Bonnefond, Mazauric, & Moutounet, 2000; Fornairon-Bonnefond 61 & Salmon, 2003); adsorbed polyphenols (Gallardo-Chacón, Vichi, Urpí, López-Tamames, & 62 Buxaderas, 2010), thiol groups of cell wall proteins (Jaehrig, Rohn, Kroh, Fleischer, & Kurz, 2007); Gallardo-Chacón et al., 2010) and β-glucans from yeast cell walls (Jaehrig et al., 2007) 63 64 also contributes to their antioxidant properties. However, ageing on the lees can modify wine 65 sensory characters, and for this reason it is not suitable for all the wine typologies; moreover, lees alone do not protect wine against microbial pollution and sulfiting is always required. 66 67 The possibility to use glutathione (GSH) as wine antioxidant has been considered since the role of this tripeptide in preventing must browning has been highlighted (Singleton, Salgues, 68 69 Zaya, & Trousdale, 1985); nevertheless, very few studies are currently available concerning 70 GSH addition in wine. It has been reported that glutathione supplementation could have 71 positive effects on wine color and aroma (Dubourdieu & Lavigne-Cruege, 2003; 72 Papadopoulou & Roussis, 2008), but high amounts of GSH in oxidative conditions can lead to 73 color formation (Sonni, Clark, Prenzler, Riponi, & Scollary, 2011). GSH can be supplemented also in form of yeast derivatives (YD): the ability of "glutathione-74 75 enriched" inactive dry yeast preparations (IDY) in reducing the loss of volatile compounds 76 during wine storage has been reported by Andújar-Ortiz, Rodríguez-Bencomo, Moreno-77 Arribas, Martin-Alvarez, & Pozo-Bayon (2010): they hypothesized that this may be due to the 78 antioxidant capacity of GSH, but also other components of the IDY preparation might be 79 involved (Andújar-Ortiz et al., 2010). 80 The opportunity to reduce sulfur dioxide, by these alternative tools, is arousing more and 81 more interest, among winemakers. Nevertheless, despite the amount of works reporting the antioxidant effects of these substances, the most of the papers regards model solutions and 82

Another traditional system to protect wine against oxidations is the use of yeast lees (Pérez-

- moreover, it is currently difficult to foresee in which extent it is possible to replace sulfites with each of these alternatives, preserving wine quality, because of the lack of scientifically-
- 85 based direct comparisons, among their effects and those of sulfur dioxide.
- 86 For this reason, the aim of this work was to carry out a preliminary investigation on the
- 87 radical scavenging activity (measured by DPPH• assay) and the oxygen consumption capacity
- 88 of different enological products and additives in comparison with SO₂. Ascorbic acid
- 89 (considered as reference standard), glutathione, yeast lees and a self-prepared yeast autolysate
- 90 were tested. Trials were performed in model solution and in different wine typologies.
- 91 Concerning oxygen consumption trials, wines were finally subjected to fast
- 92 spectrophotometric measurements, for assessing the effect of the different antioxidants on
- 93 color, total phenolics and predisposition to browning.

2 Materials and Methods

95 2.1 Chemicals

- 96 Tartaric acid, sodium hydroxide, ethanol (96 % v/v), ACS grade hydrochloric acid (37 %),
- 97 hydrogen peroxide (30 % w/w), sodium acetate and potassium metabisulfite were from Carlo
- 98 Erba Reagents (Milan, Italy); ascorbic acid, glutathione, 1,1-diphenyl-2-picryl-hydrazyl free
- 99 radical (DPPH•) and HPLC grade methanol were purchased from Sigma-Aldrich (St. Louis,
- 100 MO, USA).
- 101 2.2 Yeast lees and yeast derivatives (YD)
- Fresh lees were supplied by Viticoltori Friulani "La Delizia" (Casarsa della Delizia, PN, Italy)
- and they were obtained after alcoholic fermentation and racking of a white table wine. The
- inactive dry yeast preparation (YD) used for the trials was a thermally produced yeast
- autolysate, prepared as reported elsewhere (Comuzzo, Tat, Liessi, Brotto, Battistutta, &
- 106 Zironi, 2012).

- 107 *2.3 Evaluation of radical scavenging activity*
- 108 2.3.1 Sample preparation
- Trials were performed in model solution and wine. The former was a model buffer prepared
- by dissolving 5 g/L (33 mM) of tartaric acid in a distilled water ethanol mixture (12 % v/v);
- the pH was set at 3.20 by adding 4 M sodium hydroxide. Ascorbic acid (50 mg/L),
- glutathione (50 and 500 mg/L), potassium metabisulfite (100 and 1000 mg/L, corresponding
- respectively to 50 and 500 mg/L of sulfur dioxide), yeast lees (2.5 % v/v) and the YD
- preparation (2.5 % w/v) were added and the samples were immediately analyzed by DPPH•
- assay, as reported below.
- The wine was a white table wine from harvest 2010, supplied by Viticoltori Friulani "La
- Delizia" (Casarsa della Delizia, PN, Italy); additives, lees and YD preparation were added in
- the same amounts reported above for wine-like solution; in addition a Control sample
- 119 (untreated wine) was also included in the experimental design. Control wine and treated
- samples were subjected to DPPH• assay as reported below. All the experiments were carried
- out in three repetitions, for both wines and model solutions.
- 122 2.3.2 *DPPH•* assay
- 123 DPPH• assay was performed by a modification of the methods reported by Brand-Williams,
- 124 Cuvelier, & Berset (1995) and Gallardo-Chacón et al. (2010), using a UV-vis
- spectrophotometer (model V-530, Jasco Co. Ltd., Tokyo, Japan). A 6 × 10⁻⁵ M DPPH•
- solution was prepared fresh daily, in a 60:40 mixture of methanol : acetate buffer (0.1 M
- sodium acetate, buffered at pH 4.50 with 6 M hydrochloric acid). 3 mL of this stock solution
- were introduced in a 10 mm optical path length glass cuvette (Hellma Analytics, Mülheim,
- 129 Germany) and 100 µL of the wine samples or fresh prepared antioxidant model solutions were
- added; DPPH• discoloration was followed at 515 nm during 10 min, reading the absorbance

- against methanol: acetate buffer. Results were expressed as the percent diminution of the
- original absorbance [ΔAbs 515 nm (%)].
- For the samples treated with yeast lees and YD preparation, where insoluble particles were
- present, the reaction with DPPH• has been carried out as suggested by Gallardo-Chacón et al.
- 135 (2010): 3 mL of DPPH• and 100 µL of sample were introduced in a test tube; after 10 min,
- the reaction mixture was filtered on a 0,80 µm nylon membrane and immediately subjected to
- spectrophotometric measurement. The initial value of the absorbance was read by adding 100
- 138 µL of methanol: acetate buffer, to 3 mL of DPPH• stock solution.
- 139 Concerning model solutions, for taking into account the effect of the solvent, a blank was also
- 140 prepared, performing the DPPH• assay on the model buffer alone (tartaric acid in
- 141 hydroalcoholic solution 12 % v/v, pH 3.2); the percent values measured for the ΔAbs 515 nm
- were used to correct the analytical results.
- 143 *2.4 Oxygen consumption capacity*
- 144 *2.4.1 Equipment*
- 145 The system used for oxygen measurements was an OxySense® fluorimeter (OxySense Inc.,
- Dallas, TX, USA); O2xyDot® oxygen sensitive sensors (OxySense Inc., Dallas, TX, USA),
- were glued, by a specific silicon based oxygen permeable adhesive (OxySense Inc.), to the
- inner surface of each of the 750 mL colorless glass bottles used for the experiments. When
- O2xyDot[®] sensors are illuminated by a pulsed blue light, they emits a red fluorescent light,
- that is monitored by OxySense[®] fluorimeter. Dynamic quenching by oxygen molecules
- determines a decrease of the O2xyDot® fluorescence lifetime, that is proportional to the
- oxygen concentration in the bottles; the temperature is measured simultaneously, by an
- infrared sensor positioned in the reader pen (Li, Ashcraft, Freeman, Stewart, Jank, & Clark,
- 154 2008).

156 Two different white wines were used in two different sets of experiments. In the first one, a 157 base wine for Prosecco D.O.C.G. Conegliano Valdobbiadene (harvest 2012), supplied by 158 Cantina Produttori Valdobbiadene "Val d'Oca" (San Giovanni di Valdobbiadene, TV, Italy), 159 was used; free sulfur dioxide was 13 mg/L and alcoholic strength 10.50 % v/v. The wine 160 coming from a 20 L bulk, was shaken until oxygen saturation was reached and then transferred into the 750 mL bottles where the O2xyDot® sensors were glued (see Section 161 162 2.4.1). The average oxygen concentration measured at filling was 8.25 ± 0.19 mg/L and the 163 temperature of the samples was 19.0 °C. Ascorbic acid (50 mg/L), glutathione (50 mg/L), 164 potassium metabisulfite (100 mg/L, corresponding to 50 mg/L of sulfur dioxide), yeast lees 165 (2.0 % v/v) and YD preparation (500 mg/L) were added to the bottles; all the treatments were 166 replicated three times. Nitrogen was blown in the headspace of the samples and bottles were 167 immediately sealed with crown cap closures. A control sample (untreated wine) without any 168 antioxidant addition was also prepared in three repetitions. 169 A blend of Chardonnay, Sauvignon and Picolit from harvest 2007 (Bastianich Winery, 170 Cividale del Friuli, UD, Italy) was used for the second set of trials; free sulfur dioxide was 5 171 mg/L and alcoholic strength 13.50 % v/v. Wine was saturated with oxygen (as reported above) and then transferred in the 750 mL bottles with O2xyDot® sensor; the average oxygen 172 173 concentration at filling was 7.29 ± 0.14 mg/L and the temperature of the samples 21.0 °C. 174 Potassium metabisulfite, ascorbic acid, glutathione, YD preparation and fresh lees were added 175 to the bottles in the same amounts reported above; a control sample (untreated wine) without 176 any antioxidant addition was also prepared and three repetitions were setup for each 177 experiment. As reported for the Prosecco base wine, nitrogen was blown in the headspace of 178 the samples and bottles were immediately crown capped. For both wine typologies, oxygen 179 concentration into the bottles was measured daily and wines were stored at 20 °C during the 180 whole time of the measurements.

- 181 Spectrophotometric measures and browning assay
- 182 All the wines were analyzed 15 days after bottling; in addition, for the blended wine, analyses
- 183 were repeated after 8 months. Wine color and total phenolics were assessed by measuring the
- 184 absorbance of the samples at 420 and 280 nm respectively, using 10 mm optical path length
- 185 quartz cuvettes (Hellma Analytics, Mülheim, Germany); readings were performed against
- 186 distilled water. Concerning the UV measures, wine samples were previously diluted ten times
- 187 and total phenolic index (TPI) was calculated multiplying by 10 the absorbance measured at
- 188 280 nm.
- 189 The predisposition of wines towards browning was determined by slightly modifying the so
- 190 called POM-test, a browning test reported by Müller-Späth (1992); briefly, 5 mL of wine
- 191 were heated at 60 °C for one hour, after addition of 25 µL of a 3 % hydrogen peroxide
- 192 solution; the browning produced was estimated on the basis of the percent increase of the
- 193 absorbance at 420 nm. All the analyses were carried out by using a UV-vis
- 194 spectrophotometer, model V-530 (Jasco Co. Ltd., Tokyo, Japan).
- 195 2.5 Statistical analysis
- 196 As concerns DPPH• trials, One Way ANOVA was carried out on the percent diminutions of
- 197 absorbance, measured during the assay [Δ Abs 515 nm (%)]; means and standard deviations
- 198 (SD) were calculated and significant differences were evaluated by Tukey HSD test, at p <
- 199 0.05. Variances were homogeneous according to Brown-Forsythe test.
- 200 To assess differences in oxygen consumption rates, Factorial ANOVA was used on the
- 201 oxygen concentrations measured for the different treatments; means and standard errors (SE)
- 202 were calculated, and significant differences were assessed by Tukey HSD test (p < 0.05).
- 203 Variances were homogeneous according to Cochran C, Hartley F-max and Bartlett test.
- 204 Finally, as regards spectrophotometric measurements (Abs 280 nm, 420 nm and POM-test),
- One Way ANOVA and Tukey HSD test were carried out as reported for DPPH• assay; 205

results were considered significant at p < 0.05. All the analyses were carried out by using the software Statistica for Windows, version 8.0 (StatSoft, Inc., Tulsa, OK, USA).

3 Results and Discussion

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3.1 Evaluation of radical scavenging activity

The results of DPPH• assay are reported in Fig. 1. Concerning model solutions (Fig. 1a), the highest radical scavenging activity was detected for yeast lees, followed by YD and the highest sulfur dioxide addition (500 mg/L). At lower amounts, such as those normally used in winemaking (50 mg/L), SO₂ was less effective in bleaching the free radical and the values reported were more similar (even if statistically lower) to those measured for the same dosage of ascorbic acid (reference standard). Despite the reaction of sulfite (SO₃²-) and bisulfite ion (HSO₃⁻) with free radicals was described (Brandt & van Eldik, 1995; Neta & Huie, 1985), very few publications report data about their radical scavenging capacity in wine; moreover, the information available are generally not in agreement with the dataset presented here. In a paper published in the late 1990s, Manzocco, Mastrocola, & Nicoli (1998) found that the addition of sulfur dioxide (250 mg/L) to a model system simulating wine (water - ethanol 12 % v/v), did not affect the chainbreaking capacity of the samples towards DPPH•; the different results they obtained respect to the present study might be related to the different pH and ionic strength of the solvents used in the two experiments, for both sample preparation (hydroalcoholic solution vs. hydroalcoholic buffer, pH 3.20) and DPPH• assay (pure methanol vs. methanol: acetate buffer pH 4.50). pH might have affected the ratio between the different forms of sulfur(IV) oxides in aqueous solution and it is known that they have a different reactivity towards free radicals (Neta & Huie, 1985). Moreover, also the different ionic strength of the two model solutions might have a non-negligible role: in fact, as reported by Brandt & van Eldik

230 (1995), the reaction rate between sulfur(IV) oxides and radicals increases with the increase of 231 the ionic strength. Anyway, it is important to underline that the hydroalcoholic tartaric buffer 232 we used for dissolving or suspending the different products tested, gave a very poor effect in 233 bleaching DPPH• solution (\triangle Abs 515 nm = -3.4 %). 234 Also the results of Vivas and colleagues (Vivas, Saint-Cricq de Gaulejac, & Glories, 1997) 235 are in disagreement with the radical scavenging capacity of sulfur dioxide, reported in Fig. 1a. Studying the influence of SO₂ and ascorbic acid on the scavenging effect of tannins, they 236 237 wrote that, at usual enological concentrations, sulfites do not have a scavenging effect on 238 superoxide anion. Nevertheless, according to literature, superoxide radical reacts very slowly with sulfite (SO₃²⁻) and the rate constant of the reaction is very low (82 M⁻¹ s⁻¹) (Neta & Huie. 239 240 1985). 241 As observed for SO₂, also glutathione activity in model wine depends on the concentration. 242 GSH levels in wine range from non-detectable values to 70 mg/L (Kritzinger, Bauer, & du 243 Toit, 2013a), with a relatively high variability; at a concentration close to these amounts (50 mg/L), glutathione demonstrated the lowest ability in bleaching DPPH• and a dosage ten 244 245 times higher (500 mg/L) was needed to significantly overtake the effect of the reference 246 sample (ASC 50 mg/L); nevertheless, regardless of the amounts added, GSH activity was 247 always significantly lower than that measured for the same addition of sulfur dioxide. 248 This last considerations is not in full agreement with the data published by Vivas, Vivas de 249 Gaulejac, & Nonier (2001); they measured the scavenging activity towards superoxide anion 250 of different antioxidants in model solution, in a range of concentration from 0.1 to 1 g/L; they 251 also found that ascorbic acid was more effective than an equal amount of sulfur dioxide and 252 glutathione, but in their experiment, the scavenging activities observed for the latter additives 253 were very similar. 254 Speaking from the chemical point of view, the effects reported for such molecules in Fig. 1a seem connected quite well with their molar concentration. Glutathione molar concentration 255 11

in the sample at 50 mg/L, the one with the lowest radical scavenging activity, corresponds to approx. 0.16 mM. Ascorbic acid and sulfur dioxide at 50 mg/L, were 0.28 and 0.78 mM respectively and determined a greater percentage of discoloration; the higher efficiency of the former, despite its lower molar concentration, confirms the higher capacity of ASC to scavenge free radicals (Vivas et al., 2001). Finally, GSH at 500 mg/L (1.6 mM) and the same quantity of SO₂ (7.8 mM) showed an increasing activity in DPPH• bleaching. On the basis of this trend, a non-negligible part of the effects observable when comparing different antioxidants in winemaking conditions (therefore, in the amounts normally used and speaking in terms of mg/L) should be attributed not only to the chemical characteristics of each antioxidant molecule in itself, but also to its molar concentration. This is certainly a key point in evaluating antioxidant additives in comparison with sulfur dioxide; in fact, due to its average concentration in wine (0-40 mg/L as free SO₂) and to the lower molecular weight respect to the most suggested alternatives (ACS or GSH), the molar concentration of free SO₂ (considering both the molecular fraction and bisulfite ion) is normally higher, respect to that of these latter molecules. For instance, 240 mg/L of GSH and 137 mg/L of ascorbic acid would be required for obtaining a 0.78 mM solution, the same molar concentration of sulfur dioxide in the sample at 50 mg/L; these amounts are more than a half of the European legal limit established for ASC (250 mg/L, according to the Regulation (EC) No 606/2009) and far from the normal quantity of GSH naturally detectable in wine (Dubourdieu & Lavigne-Cruege, 2003; Kritzinger et al., 2013a; Fracassetti, Lawrence, Tredoux, Tirelli, Nieuwoudt, & du Toit, 2011; Sonni et al., 2011). Another interesting observation can be made, comparing the effects reported in Fig. 1a for the yeast autolysate (YD) and the two samples supplemented with glutathione. As reported by Kritzinger and colleagues (Kritzinger et al., 2013a), commercial YDs claim to preserve wine aroma, delaying the development of browning and oxidized notes; reduced GSH is generally considered the YD's most active component from this point of view (Pozo-Bayón,

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Andújar-Ortiz, & Moreno-Arribas, 2009; Andújar-Ortiz et al., 2010; Kritzinger et al., 2013a). According to Tirelli, Fracassetti, & De Noni (2010), glutathione content in yeast autolysates ranges from 1 to 14 mg/g (0.33-4.60 mmol/100 g), a value close to the 0.1-1 % (1-10 mg/g) of the Saccharomyces cerevisiae dry cell weight (Bachhawat et al., 2009). These quantities also agree with the information available about the ability of such products to release the tripeptide in model solution: Andújar-Ortiz and co-workers (Andújar-Ortiz, Pozo-Bayón, Moreno-Arribas, Martin-Alvarez, & Rodríguez-Bencomo, 2012) detected 1-2 mg/L of reduced GSH, after addition of 0.3 g/L of a glutathione-enriched inactive dry yeast (GSH-IDY); these data were confirmed by Kritzinger and colleagues (Kritzinger, Stander, & Du Toit, 2013b), who detected a glutathione release between 1.45 and 2.53 mg/L, after the same supplementation (0.3 g/L) with five different GSH-IDY preparations. On the basis of these quantities, we could roughly quantify in 3-8 mg the average amount of free glutathione released by 1 g of YD preparation. In the present study, we made a yeast autolysate addition of 25 g/L (2.5 % w/v), 50-100 times higher respect to the normal amounts used in winemaking (250-500 mg/L). According to the calculations above, this would be able to release approx. 75-200 mg/L of free GSH, so a lower quantity respect to the highest amount of glutathione we have added to the model wine (500 mg/L); nevertheless, the DPPH• discoloration promoted by YD in Fig. 1a is significantly higher respect to that determined by 500 mg/L of GSH. On the basis of these results, we can hypothesize that, probably, glutathione was not the only factor in determining the radical scavenging capacity of the yeast derivative, but something else among YD components might be involved, with a non-negligible contribution, in the effects observed. This hypothesis was also considered in the paper published by Andújar-Ortiz et al. (2010): reporting the effects of glutathione-enriched inactive dry yeast preparations on the aroma of wines, besides the antioxidant capacity of GSH, they also

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308 components from the IDY preparations". 309 The most accredited among these "other components" might be proteins: Jaehrig et al. (2007) 310 consider proteins from yeast cell walls as one of the most active components from the anti-311 oxidative point of view, due to their aromatic side chains and thiol groups of cysteine 312 residues. In support of this postulate, Tirelli and colleagues (Tirelli et al., 2010) quantified the 313 amounts of reducing proteins with cysteine residues (RPC), in several YD products, in 314 comparison with the content of free glutathione (GSH) and free cysteine (Cys); RPC were 315 estimated at non-negligible concentrations in the four yeast autolysates evaluated (0.73-1.40 316 mmol in 100 g of product), with values close, or even higher, respect to that of reduced GSH. 317 Obviously, in the light of these few bibliographic evidences, our current dataset must be 318 considered only as a preliminary result, being still quite poor to allow a certain conclusion; 319 further investigations will be needed to confirm these hypotheses, as well as to better 320 elucidate the role of the different YD components and fractions in scavenging free radicals. 321 A final consideration about Fig. 1a is related to the comparison between the radical 322 scavenging activity of the YD preparation and that of yeast lees. The high amount of yeast 323 autolysate used in this study was selected to make it comparable with lees addition (2.5 % 324 v/v). Obviously, we can expect that fresh lees were less concentrated in yeast cell residues, 325 because of their higher humidity content; nevertheless, the anti-radical activity of the model 326 wines treated with lees was significantly higher respect to that observed for the samples 327 treated with the yeast derivative. This could be due to the presence of residual phenolic 328 substances in the lees, that could have contributed to DPPH• discoloration, but also to the 329 production process of yeast derivatives, which could have reduced the radical scavenging 330 capacity of yeast cell components. This last observation could confirm the conclusions of 331 Tirelli et al. (2010); in the study mentioned above, they also reported the levels of overall Cys (Cys + 2 × Cystine), that is to say a parameter which also takes into account the thiol 332 14

mentioned "the activation of different types of chemical reactions promoted by other

groups in form of disulfide; this fraction was generally the most representative among those estimated (free GSH, free Cys, RPC and overall Cys), with a big variation among the analyzed YD formulations; for this reason, the authors hypothesized that the technologies applied for the production of yeast derivatives are not suitable for preserving the thiol groups of the proteins with cysteine residues, as well as the antioxidant capacity of these products. When the radical scavenging activity was measured in a white wine, the results were different respect to what observed in model solution (Fig. 1b). First of all, all the tested substances significantly increased the DPPH• bleaching capacity of the wine itself (Control sample). Lees was the most active product in promoting discoloration, but the differences among the treatments seemed less intense than those observed in the model buffer. The concentration affected in a lower extent the scavenging ability of the products: for example, the two levels tested for sulfur dioxide gave, statistically, similar performances in terms of $\Delta Abs \%$ and also the two dosages of glutathione were not as different as those reported in Fig. 1a. These behaviors might be explained considering a sort of "matrix-related effect", connected to the ability of the different antioxidants, to interact with other components present in wine. In effects, the wine itself, demonstrated a non-negligible capacity in scavenging DPPH•; the presence of the antioxidant products might have affected this capacity, not only by directly reacting with the free radical, but also by interacting with certain wine compounds. For instance, it is reported that ascorbic acid, glutathione and SO₂ can hamper oxidation chains by regenerating phenolic species from o-quinones (Waterhouse & Laurie, 2006; Danilewicz, 2007; Bradshaw et al., 2011); phenolics are well known radical scavengers and this regeneration could be connected just with the increased chain-breaking activity measured for the treated samples. It is also interesting to observe, that comparing the activity of GSH with those of ascorbic acid and SO₂ (samples at 50 mg/L), the former is no more the less effective in promoting DPPH• discoloration (as it was in model buffer): in Fig. 1b, at a low dosage, glutathione

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behaves similarly to sulfur dioxide, being not statistically different respect to the performances of ascorbic acid. In addition, at 500 mg/L, despite the five-folds lower molar concentration and in opposition to what seen in wine-like solution, GSH demonstrated a higher efficacy, respect to sulfites. Finally, a last consideration concerns the YD preparation; if compared just with glutathione (500 mg/L), one can observe that, in wine, the differences between the two treatments are basically nil. This might be explained considering the previously mentioned "matrix-related effect", so that the supplementation with pure glutathione, in a real wine, seemed to be as effective as the YD product, probably due to the capacity of the tripeptide to improve the overall radical scavenging activity of the wine itself. This might be connected with the ability of GSH to react with quinones, regenerating phenolic molecules (Waterhouse & Laurie, 2006), but further investigations shall be done to better clarify the role of glutathione and YDs in hampering radical chains in both model solution and different wine typologies.

3.2 Oxygen consumption capacity

The ability of the different antioxidant products to affect oxygen consumption was investigated in two different wines. The curves describing oxygen consumption vs. time, in a young wine (harvest 2012), are reported in Fig. 2. As one can observe, the amount of oxygen dissolved at saturation was completely depleted in the untreated wine (Control) in 192 hours (8 days). The most active oxygen scavenging substances among the tested products were fresh lees (2 % v/v) and ascorbic acid (50 mg/L). At the concentration used, pure glutathione (50 mg/L) and the yeast derivative preparation (500 mg/L) gave results similar to those observed in the Control samples. On the contrary, sulfur dioxide (50 mg/L) allowed to significantly reduce oxygen consumption, with a final average level close to 3 mg/L. In Fig. 3, the statistical confirmation of these behaviors is reported, on the basis of the results of Factorial ANOVA. Concerning SO₂, it is interesting to underline that the reduction of oxygen

depletion rate occurred in the last 48 hours of the monitoring period, while during the first six days, sulfites did not basically affect the kinetic of oxygen consumption. This behavior seems in contrast with that reported by Danilewicz and colleagues (Danilewicz et al., 2008): studying the interaction of oxygen, sulfur dioxide and 4-methylcatechol in a model wine containing iron and copper, they found that the rate of reaction of oxygen was accelerated by SO₂ addition; the explanation they gave, is that sulfites can react with quinones accelerating catechol autoxidation. They also confirmed this effect on a red wine where sulfites were eliminated by adding hydrogen peroxide; this elimination significantly reduced the ability of the wine to consume oxygen (Danilewicz et al., 2008). On the other hand, a further confirmation of these results comes also from Fig. 4, where oxygen consumption vs. time is reported for the aged wine (harvest 2007). As one can observe, fresh lees and ascorbic acid were again the most active substances in scavenging oxygen, but sulfur dioxide also increased the O₂ consumption rate; it was only a slight acceleration, but according to the results of Factorial ANOVA, oxygen depletion was significantly faster in the sulfited samples than in the Control wines (Fig. 5). On the basis of these evidences, it is difficult to explain the reason why sulfites reduced the oxygen consumption rate in the young wine (Fig. 2). This product was quite poor in phenolic compounds (the average total phenolic index of the Control samples was 3.5) and had a relatively low content of free SO₂ (see Section 2.4.2). If we consider that sulfites can accelerate catechol autoxidation, it can be hypothesized that the addition of potassium metabisulfite has actually accelerated the conversion of polyphenols into quinonic species. It has been reported that, when "4-methylcatechol is oxidized in presence of SO₂, the 38 % approximately of the quinone formed reacts with bisulfite to produce the sulfonic adduct and the most of the remainder is reduced back to catechol" (Danilewicz et al., 2008); so, we should expect a progressive reduction of the concentration of available polyphenols during the storage time. Due to the low TPI of the wine, and due to the fact that, as already mentioned,

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410 this reduction should have been faster in the sulfited samples, we can hypothesize that oxygen 411 consumption, was slowed down, in the last part of the curves of the latters (see Fig. 2), just 412 because of this faster consumption of reactive substances. 413 Another interesting observation can be done comparing the oxygen consumption curves 414 detected for the aged wine (Fig. 4) and the youngest one (Fig. 2); as one can observe, oxygen 415 disappearance was initially faster in the former, but the slope of the curves progressively 416 decreased, and after 216 hours the average level of residual oxygen (in the most of the 417 treatments) was still close to 1-1.5 mg/L; otherwise, the curves reported for the young wine 418 were less steep in their first part, but the complete oxygen consumption was obtained in less 419 than 200 hours (with the only exception of the sulfited samples). This different behavior is 420 probably related with the different phenolic content and "oxidative history" of the two wines. 421 The aged one had an average TPI of 11.6, while for the youngest one TPI was lower (only 422 3.5); the former was produced by prolonged barrel ageing, while the latter was stored in 423 stainless steel vats, so in a more protective environment with respect to oxygen contact. 424 So, the faster oxygen consumption, detected at the beginning of the monitoring period in the samples from harvest 2007, was probably due to their higher phenolic content, that might 425 426 have initially increased oxygen consumption capacity (Danilewicz et al., 2008); nevertheless, 427 the more intense contact that this wine had with oxygen during the whole production chain, 428 might have been the most important factor, responsible of the decrease of oxygen 429 consumption rate in the last part of the monitoring period. Contrary, in the youngest wine, 430 which had a lower TPI, but was probably subjected to a less intense aeration during 431 processing, oxygen depletion started slower, but it was completed in a shorter time, denoting 432 a higher capacity of such product to scavenge oxygen.

434 The effects of the different treatments on color and predisposition to browning of the young 435 wine (harvest 2012) are reported in Table 1 (Section a). As one can observe, sulfiting allowed 436 the lowest color development, followed by the treatment with YD preparation. GSH and 437 ascorbic acid gave intermediate results, while the highest color formation was detected for 438 Control samples and the wines treated with fresh lees. 439 The POM-test values confirm these behaviors: the highest levels mark out samples in which 440 phenolic fraction is more preserved, while oxidative phenomena tend to reduce the POM-test 441 index. It is interesting to underline that the lowest POM-tests were detected for the wines 442 treated with GSH and ASC, while oxidizable phenolics were better protected by sulfur 443 dioxide and YD preparation. 444 Concerning the aged wine (harvest 2007), the results were less evident (Table 1, Section b); 445 15 days after the treatments, only fresh lees gave a significant increase of the color, probably 446 due to their ability to release phenolic compounds (TPI significantly increased, according to 447 ANOVA, from a value of 11.6 \pm 0. 2 in the Control wines, to 15.0 \pm 0. 3 in the lees added 448 samples). Anyway, according to the POM-Test levels, after 15 days, potassium metabisulfite 449 and YD were the additives that allowed the best preservation of oxidizable phenolics; 450 concerning sulfites, this confirms the behaviors observed in Table 1a for the young wine, 451 while, in this case (Table 1b), the effects given by the yeast autolysate were less evident, and very close to those of glutathione and ascorbic acid. 452 453 The situation of the aged wine changed 8 months after the treatments (Table 1, Section b). 454 Sulfites demonstrated their effectiveness in protecting wine against browning, allowing only a 455 slight increase of the color during storage time. YD was the additive that behave more 456 similarly to SO₂: the color index (Abs 420 nm) of the samples treated with the autolysate 457 preparation was significantly higher than those of the sulfited wines, but significantly lower than in all the other experiments, highlighting a certain ability of such kind of preparations 10 458

459 in protecting wine against color development, over a medium-long storage time. Finally, fresh 460 lees and particularly ascorbic acid were the treatments which gave the most intense browning. 461 POM-test results confirm these considerations: SO₂ gave the highest index, followed by YD 462 and GSH, while the lowest values were detected just for ASC and fresh lees. 463 In conclusion, the products tested in this study confirmed to have different effects concerning 464 their antioxidant properties, and this reflects in a very different behavior towards the 465 modifications they can induce in wine. 466 Fresh lees were very effective in oxygen and free radical scavenging, but they increased wine 467 color by the release of phenolic compounds. Ascorbic acid confirmed its effectiveness in the 468 removal of oxygen and DPPH• free radical, but as reported elsewhere (Bradshaw et al., 2001). 469 it induced an intense browning during wine storage. Glutathione also demonstrated its ability 470 in scavenging DPPH• in wine; nevertheless, it showed a poor capacity in scavenging oxygen, 471 and its activity in protecting wine against color formation was lower than that given by 472 sulfiting (particularly after 8 months of storage). This confirms the data published by Sonni et 473 al. (2011), who reported that GSH can initially provide protection against oxidative 474 coloration, but eventually induced color formation. 475 Strictly speaking in terms of the possibility to replace sulfur dioxide, the analyses carried out 476 on wine color and predisposition to browning, highlighted that SO₂ remains the most 477 performing additive. Nevertheless, maybe surprisingly, the addition of the yeast derivative 478 preparation (YD), was the treatment that behave more similarly to sulfiting: YD was very 479 active in scavenging DPPH• free radical in both model solution and wine, and, even without 480 significantly affecting the ability of the wine to consume oxygen, it protected quite well color 481 and phenolics over a medium-length storage time (8 months). Due to the effects we have 482 observed for GSH, and the average levels reported for glutathione in inactive dry yeasts, we 483 are oriented to think that this ability of YDs might be not only connected with their capacity 484 to release the tripeptide in wine; the glutathione released might contribute, but we can 20

485 hypothesize that other components of these complex preparations might be involved in the 486 effects observed, with a non-negligible role. 487 Obviously, the bottleneck in using yeast derivatives for partially replacing sulfites in bottled 488 wines is connected with the presence of insoluble particles in the commercial preparations, 489 but this does not jeopardize the possibility to use these products during the storage in stainless 490 steel containers. From this point of view, these specific results seem interesting, because such 491 kind of preparations might allow a significant reduction of sulfite levels during wine storage, 492 with the possibility of postponing sulfiting in the production steps immediately preceding 493 bottling. Further investigations will make it possible to confirm these behaviors, clarifying the 494 mechanisms through which YDs carry out their protective action, and allowing both the 495 optimization of their use in winemaking as antioxidant formulations and the development of 496 suitable technologies for producing specific preparations for this specific winemaking use.

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References

- Andújar-Ortiz, I., Rodríguez-Bencomo, J.J., Moreno-Arribas, M.V., Martin-Alvarez, P.J.,
- & Pozo-Bayón, M.A. (2010). Role of glutathione enriched inactive yeast preparations on
- the aroma of wines. In *Proceedings of 33rd World Congress of Vine and Wine 8th*
- 506 General Assembly of the OIV (pp. 154-161). Tiblisi, Georgia.

507

502

- Andújar-Ortiz, I., Pozo-Bayon, M.A., Moreno-Arribas, M.V., Martin-Alvarez, P.J., &
- Rodríguez-Bencomo, J.J. (2012). Reversed-phase high-performance liquid₂₁

510	chromatography-fluorescence detection for the analysis of glutathione and its precursor γ -
511	glutamyl cysteine in wines and model wines supplemented with oenological inactive dry
512	yeast preparations. Food Analytical Methods, 5, 154-161.
513	
514	Bachhawat, A.K., Ganguli, D., Kaur, J., Kasturia, N., Thakur, A., Kaur, H., Kumar, A., &
515	Yadav, A. (2009). Glutathione production in yeast. In T. Satyanarayana, & G. Kunze
516	(Eds.), Yeast biotechnology: diversity and applications (pp. 259-281). Berlin: Springer.
517	
518	Bradshaw, M.P., Barril, C., Clark, A.C., Prenzler, P.D., & Scollary, G.R. (2011). Ascorbic
519	acid: a review of its chemistry and reactivity in relation to a wine environment. Critical
520	Reviews in Food Science and Nutrition, 51, 479-498.
521	
522	Bradshaw, M.P., Cheynier, V., Scollary, G.R., & Prenzler, P.D. (2003). Defining the
523	ascorbic acid crossover from anti-oxidant to pro-oxidant in a model wine matrix
524	containing (+)-catechin. Journal of Agricultural and Food Chemistry, 51, 4126-4132.
525	
526	Bradshaw, M.P., Prenzler, P.D., & Scollary, G.R. (2001). Ascorbic acid-induced
527	browning of (+)-catechin in a model wine system. Journal of Agricultural and Food
528	Chemistry, 49, 934–939.
529	
530	Brand-Williams, W., Cuvelier, M., & Berset, C. (1995). Use of a free radical method to
531	evaluate antioxidant activity. Lebensmittel-Wissenschaft und-Technologie, 28, 25-30.
532	
533	Brandt, C., & van Eldik, R. (1995). Transition metal-catalyzed oxidation of sulfur(IV)
534	oxides. Atmospheric-relevant processes and mechanisms. <i>Chemical Reviews</i> , 95, 119-190.
535	22

030	Bueuner, G., & Jurkiewicz, B.A. (1996). Chemistry and biochemistry of ascorbic acid. In
537	E. Cadenas, & L. Packer (Eds.), Handbook of Antioxidants (pp. 91-115). New York:
538	Marcel Dekker.
539	
540	Comuzzo, P., Tat, L., Liessi, A., Brotto, L., Battistutta, F., & Zironi, R. (2012). Effect of
541	different lysis treatments on the characteristics of yeast derivatives for winemaking.
542	Journal of Agricultural and Food Chemistry, 60, 3211-3222.
543	
544	Danilewicz, J.C. (2007). Interaction of sulfur dioxide, polyphenols, and oxygen in a wine-
545	model system: central role of iron and copper. American Journal of Enology and
546	<i>Viticulture</i> , 58, 53-60.
547	
548	Danilewicz, J.C. (2011). Mechanism of autoxidation of polyphenols and participation of
549	sulfite in wine: key role of iron. American Journal of Enology and Viticulture, 62, 319-
550	328.
551	
552	Danilewicz, J.C., Seccombe, J.T., & Whelan, J. (2008). Mechanism of interaction of
553	polyphenols, oxygen, and sulfur dioxide in model wine and wine. American Journal of
554	Enology and Viticulture, 59, 128-136.
555	
556	Dubourdieu, D., & Lavigne-Cruege, V. (2003). The role of glutathione on the aromatic
557	evolution of dry white wine. Wine Internet Technical Journal, 17, 1-10
558	(www.infowine.com).
559	
560	du Toit, W.J., Marais, J., Pretorius, I.S., & du Toit, M. (2006). Oxygen in must and wine:
561	a review. South African Journal of Enology and Viticulture, 27, 76-94.
	23

562	
563	Fornairon-Bonnefond, C., & Salmon, JM. (2003). Impact of oxygen consumption by
564	yeast lees on the autolysis phenomenon during simulation of wine aging on lees. Journal
565	of Agricultural and Food Chemistry, 51, 2584–2590.
566	
567	Fracassetti, D., Lawrence, N., Tredoux, A.G.J., Tirelli, A., Nieuwoudt, H.H., & du Toit,
568	W.J. (2011). Quantification of glutathione, catechin and caffeic acid in grape juice and
569	wine by a novel ultra-performance liquid chromatography method. Food Chemistry, 128,
570	1136–1142.
571	
572	Gallardo-Chacón, J.J., Vichi, S., Urpí, P., López-Tamames, E., & Buxaderas, S. (2010).
573	Antioxidant activity of lees cell surface during sparkling wine sur lie aging. International
574	Journal of Food Microbiology, 143, 48-53.
575	
576	Jaehrig, S.C., Rohn, S., Kroh, L.W., Fleischer, LG., & Kurz, T. (2007). In vitro potential
577	antioxidant activity of (1-3),(1-6)-β-D-glucan and protein fractions from Saccharomyces
578	cerevisiae cell walls. Journal of Agricultural and Food Chemistry, 55, 4710–4716.
579	
580	Kritzinger, E.C., Bauer, F.F., & du Toit, W.J. (2013a). Role of glutathione in winemaking:
581	a review. J. Agric. Food Chem., 61, 269-277.
582	
583	Kritzinger, E.C., Stander, M.A., & Du Toit, W.J. (2013b). Assessment of glutathione
584	levels in model solution and grape ferments supplemented with glutathione-enriched
585	inactive dry yeast preparations using a novel UPLC-MS/MS method. Food additives &
586	contaminants. Part A, Chemistry, Analysis, Control, Exposure & Risk Assessment, 30, 80-
587	92.

588	
589	Li, H., Ashcraft, K., Freeman, B.D., Stewart, M.E., Jank, M.K., & Clark, T.R. (2008)
590	Non-invasive headspace measurement for characterizing oxygen-scavenging in polymers
591	Polymer, 49, 4541–4545.
592	
593	Manzocco, M., Mastrocola, D., & Nicoli, M.C. (1998). Chain-breaking and oxygen
594	scavenging properties of wine as affected by some technological procedures. Food
595	Research International, 31, 673-678.
596	
597	Moreaux, V., Birlouez-Aragon, I., & Ducauze, C. (1996). Copper chelation by tryptophan
598	inhibits the copper-ascorbate oxidation of tryptophan. Redox Reports, 2, 191–197.
599	
600	Müller-Späth, H. (1992). Der POM-Test. Deutscher Weinbau, 23, 1099-1100.
601	
602	Neta, P., & Huie, R.E. (1985). Free-radical chemistry of sulfite. Environmental Health
603	Perspectives, 64, 209-217.
604	
605	Oliveira, C.M., Ferreira, A.C.S., De Freitas, V., & Silva, A.M.S. (2011). Oxidation
606	mechanisms occurring in wines. Food Research International, 44, 115-1126.
607	
608	Papadopoulou, D., & Roussis, I.G. (2008). Inhibition of the decrease of volatile esters and
609	terpenes during storage of a white wine and a model wine medium by glutathione and N-
610	acetylcysteine. International Journal of Food Science and Technology, 43, 1053–1057.
611	
612	Pérez-Serradilla, J.A., & Luque de Castro, M.D. (2008). Role of lees in wine production: a

review. Food Chemistry, 111, 447–456.

614 Pozo-Bayón, M.Á., Andújar-Ortiz, I., & Moreno-Arribas, M.V. (2009). Scientific 615 616 evidences beyond the application of inactive dry yeast preparations in winemaking. Food 617 Research International, 42, 754-761. 618 619 Ribéreau-Gayon, P., Dubourdieu, D., Doneche, B., & Lonvaud, A. (2006). Handbook of Enology. Vol. 1. The Microbiology of Wine and Vinifications. (2nd ed.). New York: John 620 621 Wiley & Sons. 622 623 Salmon, J.-M., Fornairon-Bonnefond, C., Mazauric, J.-P., & Moutounet, M. (2000). 624 Oxygen consumption by wine lees: impact on lees integrity during wine ageing. Food 625 Chemistry, 71, 519-528. 626 627 Singleton, V.L. (1987). Oxygen with phenols and related reactions in musts, wines, and 628 model systems: observations and practical implications. American Journal of Enology and Viticulture, 38, 69-77. 629 630 Singleton, V.L., Salgues, M., Zaya, J., & Trousdale, E. (1985). Caftaric acid 631 632 disappearance and conversion to products of enzymic oxidation in grape must and wine. American Journal of Enology and Viticulture, 36, 50-56. 633 634 635 Sonni, F., Clark, A.C., Prenzler, P.D., Riponi, C., & Scollary, J.R. (2011). Antioxidant 636 action of glutathione and the ascorbic acid/glutathione pair in a model white wine. Journal 637 of Agricultural and Food Chemistry, 59, 3940–3949.

039	Tirein, A., Fracassetti, D., & De Noni, I. (2010). Determination of reduced cysteine in
640	oenological cell wall fractions of Saccharomyces cerevisiae. Journal of Agricultural and
641	Food Chemistry, 58, 4565-4570.
642	
643	Vivas, N., Saint-Cricq de Gaulejac, N., & Glories, Y. (1997). Influence de SO ₂ et de
644	l'acide ascorbique sur l'activité antiradicalaire des tanins, mesurée sur l'anion superoxyde.
645	Application aux vins rouges. Vitis, 36, 91-96.
646	
647	Vivas, N., Vivas de Gaulejac, N., & Nonier, M.F. (2001). Scavenging and antioxidant
648	properties of different soluble compounds in hydroalcoholic media. Comparison with SO ₂
649	efficiency. Bulletin de l'O.I.V., 843-844, 331-346.
650	
651	Waterhouse, A.L., & Laurie, V.F. (2006). Oxidation of wine phenolics: a critical
652	evaluation and hypotheses. American Journal of Enology and Viticulture, 57, 306-313.
653	
654	Zoecklein, B.W., Fugelsang, K.C., Gump, B.H., & Nury, F.S. (1995). Sulfur Dioxide and
655	Ascorbic Acid. In, Wine Analysis and Production (pp. 178-191). New York: Chapman
656	and Hall.

Figure Captions 657 658 Fig. 1. Radical scavenging activity of different enological additives and products in model solution 659 (a) and in white table wine (b). Results of ANOVA analysis and Tukey HSD test; means and 660 standard deviations of three repetitions are reported. Different letters mark significant differences at 661 p < 0.05. See the text for abbreviations. 662 663 Fig. 2. Oxygen consumption versus time, measured for the samples prepared in Prosecco base wine 664 (harvest 2012). Mean values of three repetitions are reported. Vertical bars represent standard deviations. See the text for abbreviations. 665 666 667 Fig. 3. Factorial ANOVA and Tukey HSD test, carried out on the oxygen concentrations measured for the curves in Fig. 2. Means and standard errors (SE) of three repetitions are reported; different 668 669 letters mark significant differences at p < 0.05. See the text for abbreviations. 670 671 Fig. 4. Oxygen consumption versus time, measured for the samples prepared in the blended wine from harvest 2007 (Chardonnay, Sauvignon, Picolit). Mean values of three repetitions are reported. 672 673 Vertical bars represent standard deviations. See the text for abbreviations. 674 Fig. 5. Factorial ANOVA and Tukey HSD test, carried out on the oxygen concentrations measured 675 for the curves in Fig. 4. Means and standard errors (SE) of three repetitions are reported; different 676 677 letters mark significant differences at p < 0.05. See the text for abbreviations. 678 679 680

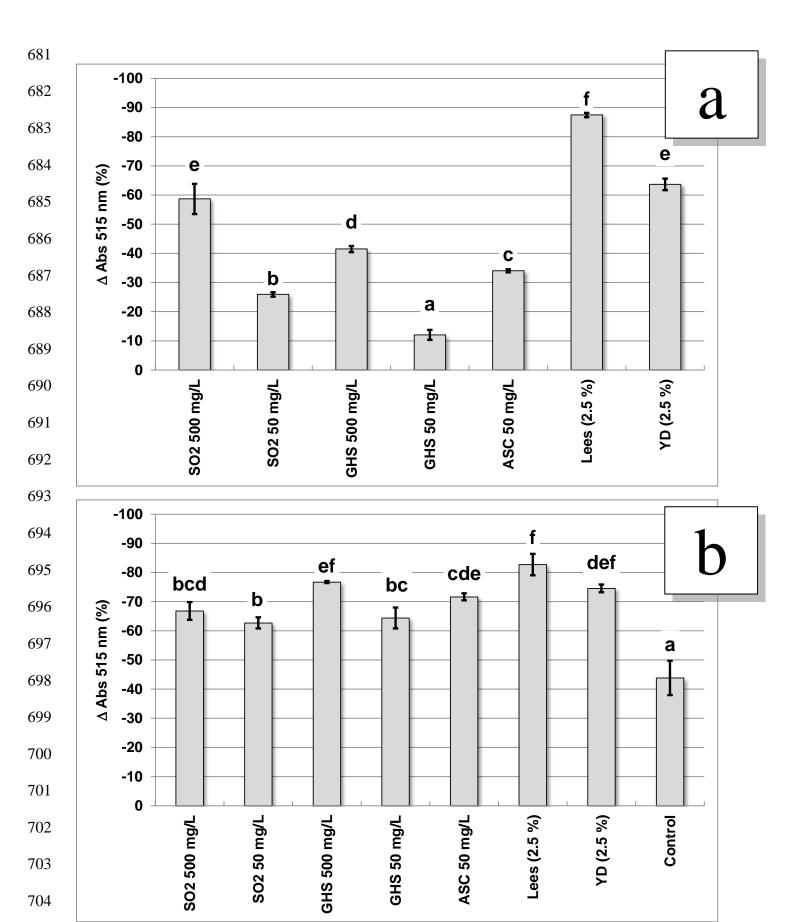


Fig. 1

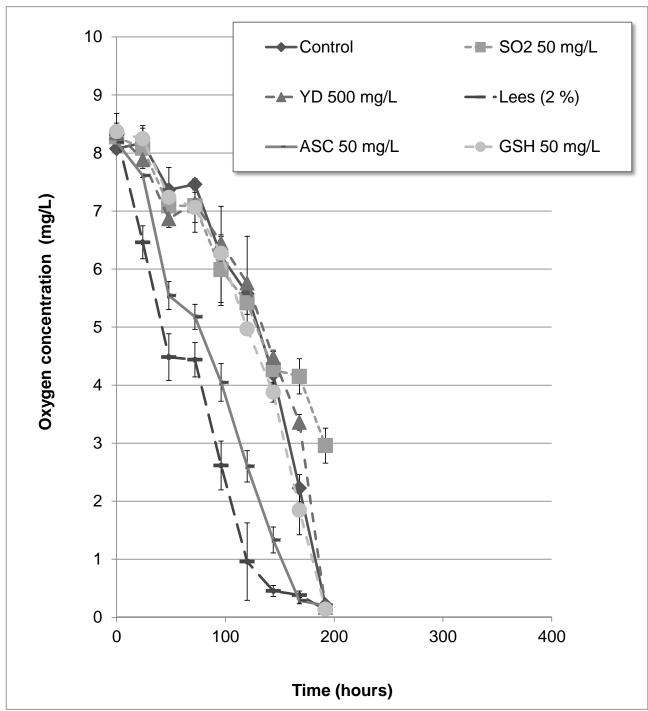


Fig. 2

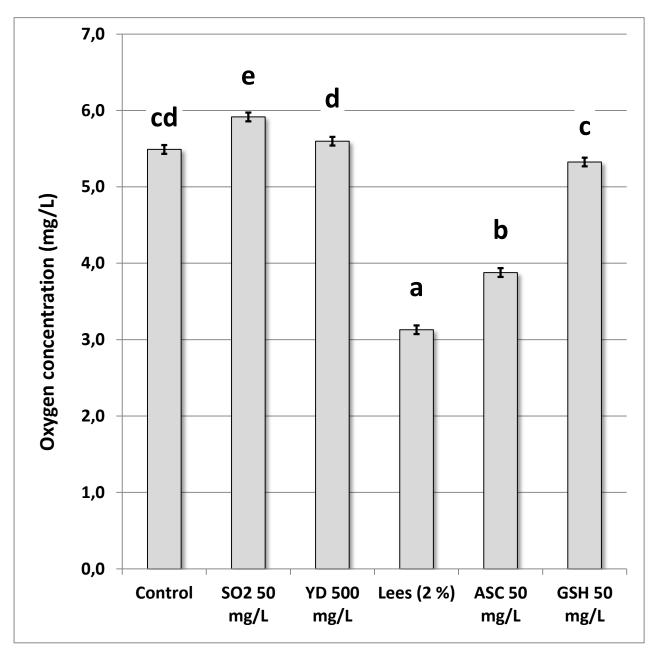


Fig. 3

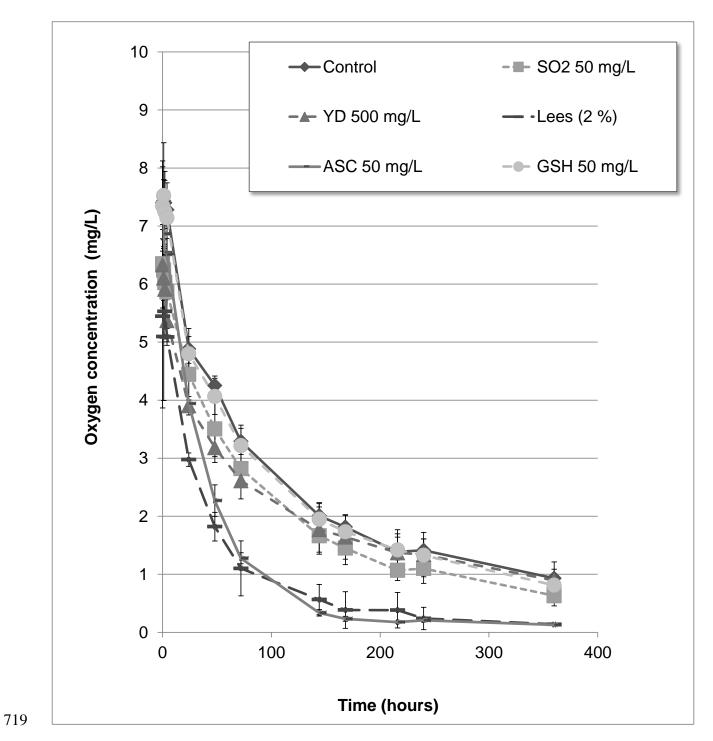


Fig. 4

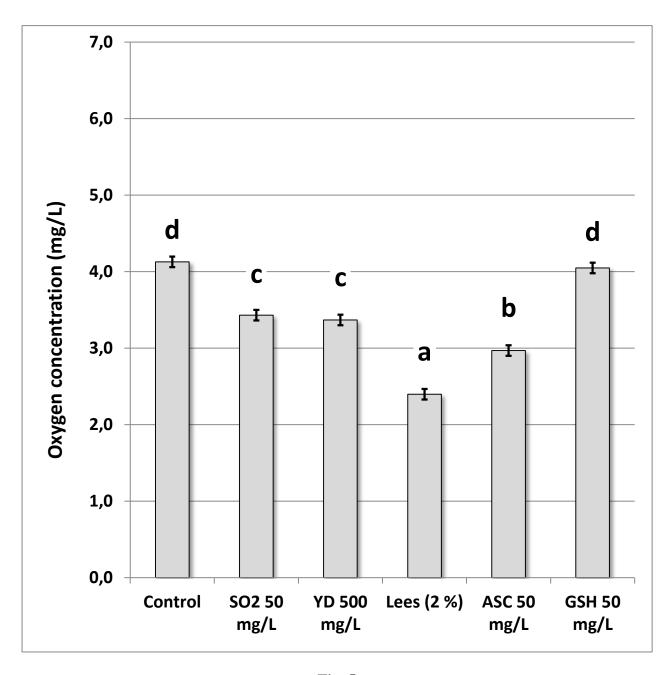


Fig. 5

Table 1

Absorbance at 420 nm and POM-test values detected for the two wines used in the experiments; samples were analyzed 15 days after bottling, and limitedly to the aged wine (harvest 2007), after 8 months. Means, standard deviations (SD) and the results of One Way ANOVA and Tukey HSD test are reported; different letters represent significant differences at p < 0.05.

(a) Young wine (harvest 2012)				
	Abs 420 nm			
Sample	15 days			
	Mean ± SD			
Control	0.038 ± 0.001 e			
SO2 50 mg/L	0.019 ± 0.000 a			
YD 500 mg/L	$0.024 \ \pm \ 0.000 b$			
Lees (2 %)	0.039 ± 0.000 e			
ASC 50 mg/L	0.033 ± 0.001 d			
GSH 50 mg/L	$0.028 ~\pm~ 0.000 ~~c$			

(b) Aged wine (harvest 2007)				
	Abs 420 nm			
Sample	15 days		8 months	
	Mean ± SD		Mean ± SD	
Control	0.204 ± 0.002	a	0.251 ± 0.005	c
SO2 50 mg/L	0.182 ± 0.002	a	0.200 ± 0.000	a
YD 500 mg/L	0.206 ± 0.024	a	0.214 ± 0.009	b
Lees (2 %)	0.273 ± 0.010	b	0.280 ± 0.005	d
ASC 50 mg/L	0.198 ± 0.001	a	0.288 ± 0.001	d
GSH 50 mg/L	0.205 ± 0.001	a	0.258 ± 0.005	c

	POM-test	
Sample	15 days	
	Mean ± SD	_
Control	90 ± 10	b
SO2 50 mg/L	$167 ~\pm~ 36$	cd
YD 500 mg/L	181 ± 6	d
Lees (2 %)	111 ± 17	bc
ASC 50 mg/L	20 ± 35	a
GSH 50 mg/L	25 ± 2	a

	POM-test			
Sample	15 days		8 months	
	Mean ± SD		Mean ± SD	
Control	35 ± 15	a	24 ± 2	bc
SO2 50 mg/L	69 ± 5	b	45 ± 0	e
YD 500 mg/L	43 ± 14	ab	29 ± 3	d
Lees (2 %)	30 ± 9	a	21 ± 1	ab
ASC 50 mg/L	42 ± 1	a	19 ± 0	a
GSH 50 mg/L	42 ± 5	a	27 ± 1	cd

*Highlights (for review)

Highlights

Sulfur dioxide was the most active additive in reducing wine color development Ascorbic acid was a good O_2 and radical scavenger but induced browning during storage Lees were good O_2 and radical scavengers but increased color by releasing phenolics Glutathione had radical scavenging ability in wine but increased color during storage Yeast derivative behave more similarly to SO_2 protecting wine color over 8 months