

Special Report

Ten Years of VINQUEST: First Insight for Breeding New Apple Cultivars With Durable Apple Scab Resistance

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Abstract

Apple scab, caused by *Venturia inaequalis*, is a major fungal disease worldwide. Cultivation of scab-resistant cultivars would reduce the chemical footprint of apple production. However, new apple cultivars carrying durable resistances should be developed to prevent or at least slow the breakdown of resistance against races of *V. inaequalis*. One way to achieve durable resistance is to pyramid multiple scab resistance genes in a cultivar. The choice of the resistance genes to be combined in the pyramids should take into account the frequency of resistance breakdown and the geographical distribution of apple scab isolates able to cause such breakdowns. In order to acquire this information and to make it available to apple breeders, the VINQUEST project (www.vinquest.ch) was initiated in 2009. Ten years after launching this project, 24 partners

from 14 countries regularly contribute data. From 2009 to 2018, nearly 9,000 data points have been collected. This information has been used to identify the most promising apple scab resistance genes for developing cultivars with durable resistance, which to date are: *Rvi5*, *Rvi11*, *Rvi12*, *Rvi14*, and *Rvi15*. As expected, *Rvi1*, together with *Rvi3* and *Rvi8*, were often overcome, and have little value for scab resistance breeding. *Rvi10* may also belong to this group. On the other hand, *Rvi2*, *Rvi4*, *Rvi6*, *Rvi7*, *Rvi9*, and *Rvi13* are still useful for breeding, but their use is recommended only in extended pyramids of ≥ 3 resistance genes.

Keywords: *Venturia inaequalis*, apple breeding, apple scab, durable resistance, molecular marker, virulence

Apple scab, caused by the ascomycete fungus *Venturia inaequalis*, is a major fungal disease in most temperate growing regions around the world, requiring many fungicide applications. Generally, up to 15

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applications of fungicides are necessary to control the disease (MacHardy 1996). However, in some areas with high precipitation and high disease pressure, 20 to 30 applications may be required when growing susceptible cultivars (Manktelow et al. 1996), thereby increasing the risk of fungicide breakdown (Ayer et al. 2019; Chapman et al. 2011; Frederick et al. 2014). This problem may have been further exacerbated through integrated pest management strategies aimed at reducing fungicide inputs for disease control (Beckerman et al. 2015). While some efficacy may still be achieved for these fungicides (van den Bosch et al. 2018), application of complementary strategies for disease control, such as orchard sanitation measures,

is essential. Thus, leaf litter management (Holb 2006, 2007; Mac an tSaoir et al. 2010; Porsche et al. 2017) assists in reducing the disease pressure and the risk of fungicide breakdown. Growing scab-resistant cultivars could be an additional contributor to more sustainable apple production, provided that the resistance is durable. Unfortunately, most of the scab-resistant cultivars released to date carry only one gene, *Rvi6* (*Vf*), and its value is questionable since *avrRvi6* isolates now occur all over Europe (Bengtsson et al. 2000; Lefrancq and Lateur 2009; Parisi et al. 2004) and have also been observed in the U.S.A. (Beckerman et al. 2009; Papp et al. 2020). Isolates virulent to other *Rvis* were identified early on in the Purdue-Rutgers-Illinois back-cross program for *Rvi2* (*Vh2*) and *Rvi4* (*Vh4*) (Williams and Shay 1957) and *Rvi5* (*Vm*) (Williams and Brown 1968). Today, virulent isolates have been shown to exist for most of the scab resistance genes used in apple breeding, including some carrying multiple virulence (Bus et al. 2011; Caffier et al. 2015; Peil et al. 2018). These findings accentuate the need for breeding for durable resistance.

A common strategy to achieve durable resistance is through pyramiding resistance genes, augmented by marker-assisted selection (MAS) technology (Vanderzande et al. 2018). The availability of a diverse range of resistance genes, enabling multiple combinations, greatly assists in the successful application of this strategy. To date, many scab resistance genes are known, of which 19 have been included in the *Rvi* nomenclature system (Bus et al. 2011; Gardiner et al. 2018; Soriano et al. 2014). Therefore, genetic markers for most of these genes, preferably cheap high-throughput single nucleotide polymorphism (SNP) and competitive allele-specific PCR (CASP) markers (Baumgartner et al. 2016; Jansch et al. 2015), are available for MAS. Also, breeding parents carrying resistance genes in a homozygous state, e.g., *Rvi2*, *Rvi4*, and *Rvi6*, have been developed (Baumgartner et al. 2015), which increases breeding efficiency and reduces the need for MAS as 100% of the progeny are resistant.

Hitherto not utilized resistance genes are available in wild apple germplasm. Fast-breeding technologies may be required to rapidly transfer these into elite germplasm, e.g., physiological (Volz et al. 2009) and/or genetic modification approaches, such as early flowering (Flachowsky et al. 2011; Schlathöler et al. 2018). In the U.S.A., null segregants (i.e., the final products of the early flowering system that do not carry the fast-flowering gene) are exempted from regulations concerning genetically modified organisms, as long as these genotypes have been shown, with molecular methods, to not contain transgenes or pieces of transgenes (Firko 2014; Gregoire 2011; McGarry et al. 2017).

With markers available for a range of resistance genes and the ability to create new cultivars with gene pyramids, the next step is to identify the most desirable gene combinations for different geographic regions. The VINQUEST project was initiated 10 years ago (Patocchi et al. 2009; www.vinquest.ch) to collect information on the geographical distribution of (a)virulence genes in *V. inaequalis* populations, following in the tradition of virulence monitoring in, for example, rust diseases in cereals (Kosman et al. 2019; Paczós-Grzęda and Sowa 2019). The VINQUEST project takes a systematic worldwide approach, extending on the standalone monitoring previously performed in a number of locations (e.g., Bengtsson et al. 2000; Lefrancq and Lateur 2009; Lespinasse 1989; Sandskär and Liljeroth 2005). A set of differential apple hosts carrying a range of scab resistance genes was developed based on the work of Bus et al. (2011) and distributed to partners around the world. In this paper, we report on the observations made at 24 trap orchards to date.

Materials and Methods

The 16 differential *Malus* hosts, representing a range of apple scab resistance genes including one host, h(0), not carrying any scab resistance genes of breeding value, had been selected previously (Table 1; Patocchi et al. 2009). Budwood of these genotypes was provided free of charge to the project participants. Each partner was free to design their own experimental orchard, but the planting of five blocks, each containing one tree of each differential host as well as five “placeholders” in a randomized fashion, was advised. The placeholders are apple scab-susceptible genotypes to be substituted once new

differential hosts become available. At some sites, few differential hosts were missing because of unsuccessful grafting. This was the case for 11 differential hosts (Table 1), but in particular for host (11), *M. baccata* ‘Jackii,’ which is present in only 17 out of the 24 possible sites. It was observed that budding of h(11) in late summer leads to a higher success rate compared with bench grafting in winter.

No fungicides were applied at any time before scoring. Afterward, application of fungicides was allowed if needed in order to protect the trees from too much damage, which might jeopardize their survival and growth until the following season.

Incidence and severity of scab on the leaves of each tree was scored on a 1 to 9 scale at the end of each apple scab primary season (Patocchi et al. 2009; Fig. 1). Only strongly sporulating lesions without resistance reactions by the host observed on young leaves (i.e., fully compatible interaction) were considered as a resistance gene breakdown. Resistance gene breakdown was documented with digital photographs. Scoring values, photographs, and in a few cases, affected leaves were sent to the coordinator (Agroscope) for validation of the observations. If doubts remained over the resistance gene breakdown being complete, the “*in dubio pro reo*” rule was applied in favor of the host; doubtful lesions were not considered as a compatible interaction and the calls were revised to disease severity score 1 (i.e., no symptoms).

An observation was defined as the score of a tree in a location (site) in a specific year. If all the observations in a specific year and locations were equal to “1”, i.e., no scab found, none of the observations of this location and year were considered for this study, due to uncondusive weather conditions for scab infection. This occurred only three times: Ora (Italy) in 2017 and 2018 and Kristianstad (Sweden) in 2016. *Rvi* genes were divided into categories based on the frequency of their breakdown over the total number of observations, or the frequency of sites in which an *Rvi* breakdown was observed at least once over the total number of sites in which the corresponding differential host had been planted. The categories were defined as follows: “often overcome,” breakdown observed in more than 50% of the observations or sites; “frequently overcome,” breakdown observed in 31 to 50% of the observations or sites; “sometimes overcome,” breakdown observed in 16 to 30% of the observations or sites; “rarely overcome,” breakdown observed in up to 15% of the observations or sites; “not overcome,” breakdown not observed. The same categories were used independently of the threshold set to call an *Rvi* gene breakdown (i.e., scores ≥ 2 or > 2).

Results

Since the start of VINQUEST 10 years ago, 24 partners from 14 countries have joined the monitoring of apple scab resistance gene breakdown and provide data (see affiliation list of the coauthors of this paper). Another 15 partners in an additional nine countries are presently establishing their differential orchards, while nine more partners in a further three countries have declared an interest in joining the project, but for different reasons have not yet done so (see www.vinquest.ch).

To date, nearly 9,000 data points have been collected (Fig. 1) and these were used to extract the first trends. Since the determination of a resistance gene breakdown is not always clear-cut, we used several approaches to interpret the collected data. In the first approach, all data were considered independently from observation site. Based on the number of disease severity scores ≥ 2 as a ratio of the total number of observations for each differential host, the hosts were assigned to a frequency category. Scab resistance genes *Rvi1*, *Rvi3*, and *Rvi8* fell in the category “often overcome”, and *Rvi6*, *Rvi7*, and *Rvi10* in the category “sometimes overcome”. Seven genes, *Rvi2*, *Rvi4*, *Rvi5*, *Rvi9*, *Rvi12*, *Rvi13*, and *Rvi14*, were assigned to the category “rarely overcome,” while *Rvi11* and *Rvi15* were assigned to the category “not overcome” since no compatible interactions were observed on their hosts to date (Table 1).

Our second approach to interpret the data without taking sites into consideration was to place the threshold to call an *Rvi* gene breakdown a disease severity class 3 score. Class 3 scores are defined as “Immediately apparent lesions generally clustered in a few parts of

the tree (1–5% of leaves affected).” Starting from this class, the scab symptoms are generally obvious. Using this threshold, *Rvi1* remains in the category “often overcome,” while *Rvi3* and *Rvi8* move to the category “frequently overcome.” Genes *Rvi6*, *Rvi7*, and *Rvi10* also change category and are now together with *Rvi2*, *Rvi4*, *Rvi5*, *Rvi9*, *Rvi12*, *Rvi13*, and *Rvi14* in the category “rarely overcome.” Obviously, *Rvi11* and *Rvi15* stay in category “not overcome” (Table 1).

Our third approach was to quantify the frequency of *Rvi* breakdowns over the different sites (Table 1). We considered an *Rvi* gene to be overcome at a specific site each time at least one class 3 score was called. This approach grouped the *Rvi* genes as follows: *Rvi1*, *Rvi3*, *Rvi8*, and *Rvi10* fell in the category “often overcome;” *Rvi2*, *Rvi4*, *Rvi6*, *Rvi7*, *Rvi9*, and *Rvi13* in “sometimes overcome;” *Rvi5*, *Rvi12*, and *Rvi14* in “rarely overcome,” and again *Rvi11* and *Rvi15* in the category “not overcome.”

The fourth approach combines the frequency of scores >2 with the frequency with which these scores had been given in the different sites (Table 1). For *Rvi1*, *Rvi3*, and *Rvi8*, observations of scores >2 occurred often or frequently, and these were also found in many of the sites. *Rvi10* rarely obtained scores >2, but such scores were nevertheless present in about half of the sites (category “often”). A relatively large group of genes, i.e., *Rvi2*, *Rvi4*, *Rvi6*, *Rvi7*, *Rvi9*, and *Rvi13*, were rarely scored >2, but such scores were still given in about a quarter of the sites (category “sometimes”). Finally, *Rvi5*, *Rvi12*, and *Rvi14* rarely received scores >2 at any site (category “rarely”).

Update on the differential hosts. The VINQUEST project is a work in progress as new differential hosts are identified following genetic studies on new scab resistance genes. Here we present an update for a number of scab resistance genes and their representative differential hosts, three of which are currently being multiplied for distribution among the VINQUEST participants.

Host (3). The current differential h(3) accession Q71 was selected from a ‘Geneva’ × ‘Braeburn’ family based on the assumption that it carried one of the two or three genes described in early reports on the ‘Geneva’ scab resistance (Bus et al. 2011). Later, this resistance was shown to be more complex and to contain a cluster of five genes, each with a moderate resistance spectrum, mapping to linkage group 4 (LG4) of this cultivar (Bastiaanse et al. 2016). Also, ‘Geneva’ itself has been reported to show scab symptoms (Beckerman et al. 2009; Bengtsson et al. 2000; Sandskär and Liljeroth 2005), although it performed well in a long-term Belgian study (Lefrancq and Lateur 2009). To determine whether this is the case for other sites as well,

we will revert to using ‘Geneva’ as h(3) (Lespinasse 1994) in *V. inaequalis* monitoring for subsequent years, while avoiding overloading the trap orchards with differential hosts representing its individual genes that on their own have little value for breeding.

Host (7). To date, in the absence of a specific h(7) only carrying the *Rvi7* gene, host (6,7) *Malus floribunda* 821 was used as it is the original source of the gene (Bénaouf and Parisi 1997, 2000). Following the mapping of the gene to LG8 of *M. floribunda* 821 (Durel et al. 2006), progeny LPG3-29 was selected as h(7). In a study with a limited number of isolates, nine out of 21 isolates were compatible with h(7) (Caffier et al. 2015), while *M. floribunda* 821 was found infected in a number of studies (Bengtsson et al. 2000; Lefrancq and Lateur 2009; Roberts and Crute 1994; Sandskär and Liljeroth 2005; this study). Like ‘Geneva,’ LPG3-29 will be monitored for a number of years in order to assess its breeding value.

Host (9). Initially, the host-pathogen interaction of *V. inaequalis* race (1,2,8,9) isolate 1639 with progeny J34 selected from a ‘Gala’ × ‘Dolgo’ family, was rated as compatible (Broggini et al. 2011) and interpreted as confirming the ability of this isolate to overcome *Rvi9*. However, later reinterpretation concluded that the interaction was incompatible, with a weak, sporulating resistance reaction suggesting the presence of a partial resistance gene. This was further substantiated when *Vdg2* was shown to segregate as a single gene in a ‘Golden Delicious’ × K108 progeny inoculated with *V. inaequalis* isolate 1639 (which is compatible with *Rvi1* [Caffier et al. 2015]). In contrast, 1639 showed a virulent phenotype with unlimited sporulation (compatible interaction) on accession K2 as well as on nearly all ‘Gala’ × K2 progeny, confirming K2 to be the preferred host (9) (V. G. M. Bus, unpublished data). An additional practical reason to change the genotype representing h(9) is that J34 has been growing poorly at several sites. The leaves often are chlorotic and fold along the main vein, making scab assessment difficult.

Host (16). The scab resistance of the open-pollinated MIS (mildew immune seedling) derivative 93.051 G07-098 was mapped as a single gene, *Rvi16*, to LG3 (Bus et al. 2010). With the aid of the microsatellite markers NH030a and NZmsCN943818, it was shown that the gene originated from MIS. However, further genetic studies with the open-pollinated MIS derivative since indicated that it contains at least one more scab resistance gene from either MIS or the unknown pollen parent. While awaiting completion of the genetic studies, MIS itself will be added to the differential host set for distribution to the trap orchards.

Host (17). Multiple studies over the years have shown that the ‘Antonovka’ scab resistance is complex, with a number of its

Table 1. Differential apple hosts carrying specific *Rvi* scab resistance genes ranked according to the percentage of resistance breakdown (class ≥2) and the frequency of such events over the sites that have provided data to date

Differential host	Genotype	<i>Rvi</i> gene	Old name	Total observations	Total obs. score ≥2 (%)	Category ^a	Total obs. score >2 (%)	Category ^a
h(0)	Gala	None	-	601	540 (89.9)	often	476 (79.27)	often
h(1)	Golden Delicious	<i>Rvi1</i>	<i>Vg</i>	609	550 (90.3)	often	488 (80.1)	often
h(3)	Q71	<i>Rvi3</i>	<i>Vh3.1</i>	582	344 (59.1)	often	252 (43.3)	frequently
h(8)	B45	<i>Rvi8</i>	<i>Vh8</i>	603	388 (64.3)	often	268 (44.4)	frequently
h(10)	A723-6	<i>Rvi10</i>	<i>Va</i>	574	159 (27.7)	sometimes	67 (11.7)	rarely
h(6)	Priscilla	<i>Rvi6</i>	<i>Vf</i>	575	95 (16.5)	sometimes	69 (12.0)	rarely
h(6,7)	<i>M. floribunda</i> 821	<i>Rvi6</i> , <i>Rvi7</i>	<i>Vf</i> , <i>Vfh</i>	515	98 (19.0)	sometimes	74 (14.4)	rarely
h(13)	Durello di Forlì	<i>Rvi13</i>	<i>Vd</i>	495	76 (15.4)	rarely	25 (5.1)	rarely
h(2)	TSR34T15	<i>Rvi2</i>	<i>Vh2</i>	537	62 (11.5)	rarely	23 (4.3)	rarely
h(4)	TSR33T239	<i>Rvi4</i>	<i>Vh4</i>	567	27 (4.8)	rarely	14 (2.5)	rarely
h(9)	J34	<i>Rvi9</i>	<i>Vdg</i>	602	49 (8.1)	rarely	14 (2.3)	rarely
h(14)	Dülmener Rosenapfel	<i>Rvi14</i>	<i>Vdr1</i>	565	16 (2.8)	rarely	3 (0.5)	rarely
h(5)	9-AR2T196	<i>Rvi5</i>	<i>Vm</i>	551	27 (4.9)	rarely	11 (2.0)	rarely
h(12)	Hansen’s baccata 2	<i>Rvi12</i>	<i>Vb</i>	569	3 (0.5)	rarely	1 (0.2)	rarely
h(11)	<i>M. baccata</i> ‘Jackii’	<i>Rvi11</i>	<i>Vbj</i>	287	0	not overcome	0 (0)	not overcome
h(15)	GMAL2473	<i>Rvi15</i>	<i>Vr2</i>	582	0	not overcome	0 (0)	not overcome

(Continued on next page)

^a Often overcome: breakdown (class ≥2 or >2) observed on more than 50% of the observations or sites; frequently overcome: breakdown (class ≥2 or >2) observed between 50 and 31% of the observations or sites; sometimes overcome: breakdown (class ≥2 or >2) observed between 30 and 16% of the observations or sites; rarely overcome: breakdown (class ≥2 or >2) observed between 1 and 15% of the observations or sites; not overcome: breakdown not observed.

progeny that have been used as breeding parents carrying different components of the resistance (Bus et al. 2012). The parents used in many breeding programs appear to have derived from ‘Common Antonovka,’ also known as ‘Schmidt’s Antonovka’ (Pikunova et al. 2014), since Schmidt widely distributed progeny from his genetic studies on scab resistance (Bus et al. 2012). The progeny mostly showed a hypersensitive response (HR) or chlorotic (Chl) resistance reactions (Brauns 1962). Currently in VINQUEST, the gene conditioning HR is associated with differential host (10), represented by accession A723-6 derived from the original *Rvi10* source PI 172623 (Bus et al. 2012). This may be the same gene as the HR-conditioning *Va2* gene mapped to a similar region, about 14 cM above the *Rvi6* region on LG1 in ‘Antonovka’ APF22 (Dunemann and Egerer 2010). The second, Chl-conditioning gene from ‘Antonovka’ APF22 was mapped to the *Rvi6* region but shown to be different from *Rvi6*. The gene was named *Rvi17* (Dunemann and Egerer 2010) and recently the genotype 04/214-079 from the ‘Golden Delicious’ × ‘Antonovka’ APF22 family was made available for distribution in the VINQUEST project. Nevertheless, further research is required to elucidate the genetic relationships of the *Rvi10* and *Rvi17* genes with those in other related ‘Common Antonovka’ derivatives, such as CCR1T8 and CCR3T11 (Lespinasse 1989), ‘Freedom’ (Zini 2005), and TN10-8 (Calenge et al. 2004).

Host (18). *Rvi18* has been identified in genotype 1980-015-025 (Soriano et al. 2014). Since it also carries the narrow spectrum *Vd3* scab resistance gene (Soriano et al. 2009), a progeny of 1980-015-025 only carrying *Rvi18* would be the natural differential host for this gene. Since no genotype will be made available unconditionally at this stage, we refrain from including h(18) trees in the trap orchard.

Hosts (19) and (20). Anticipating broad-spectrum genes to control scab resistance in ‘Honeycrisp,’ the denotations *Rvi19* and *Rvi20* were initially assigned to the two scab resistance genes identified in this cultivar (Clark 2014; Clark et al. 2014), and repeated in a number of publications (e.g., Igarashi et al. 2016; Khajuria et al. 2018). However, the *Rvi* assignments turned out to have been premature as more information became available. With the gene identified as *Rvi19* mapping to the same region on LG1 as the Chl-conditioning ‘Common Antonovka’ and showing the same CH-Vf1-139 bp allele (Clark 2014), their genetic relationships need to be elucidated. The gene identified as *Rvi20* and mapping to LG15 of ‘Honeycrisp’ appears to have a narrow spectrum, hence its breeding value is too low for inclusion in the trap orchard evaluation network. Henceforth,

the genes have been assigned *Vhc1* and *Vhc15* as working names, with *hc* referring to ‘Honeycrisp’ and the numbers to the LG they map. In the meantime, *Rvi19* has now been assigned to a third scab resistance gene from Russian apple R12740-7A (Gardiner et al. 2018). Differential hosts for *Rvi19* and *Rvi20* will be included in the *V. inaequalis* population monitoring in due course.

Discussion

The VINQUEST initiative has started well and the network continues to expand. Over the first 10 years of *Rvi* monitoring activities, sufficient data have been collected to identify the first trends. Different interpretations of the collected data have been presented, setting different thresholds to call a resistance gene breakdown; i.e., severity scores (sometimes in the text also called classes for simplicity) ≥ 2 or >2 , and grouping the *Rvis* in categories.

Our first approach defined the threshold to call an *Rvi* breakdown as starting from the identification of a few sporulating apple scab lesions without detection of any resistance reaction upon scrutiny of the tree (severity class 2). This class is the most prone to false-positive errors. While inspecting a tree, necrotic flecks on leaves may be erroneously interpreted as old apple scab lesions instead of being recognized as an expression of partial resistance. Such flecks may also be due to other fungi. Due to the awareness of these possible sources of mistakes, class 2 scores were carefully scrutinized during the validation process of checking the photographs and/or the leaves provided. When doubts remained about putatively affected leaves, the class 2 calls were revised to class 1 (application of the *in dubio pro reo* rule) to minimize the risk of calling false resistance breakdowns, so as not to discourage apple breeders from investing in some very valuable resistance sources.

In order to reduce the risk of false calls, we propose considering a resistance gene breakdown from class 3 only. Starting from this class, apple scab symptoms are sufficiently abundant and obvious, so fewer mistakes can be expected. The change of threshold from class 2 to class 3 resulted in a shift of some *Rvi* genes to lower categories; two of them (*Rvi3* and *Rvi8*) moved into the category “frequently overcome,” while another four (*Rvi6*, *Rvi7*, *Rvi10*, and *Rvi13*) moved into the category “rarely overcome.” *Rvi3* and *Rvi8* still remain in a category with quite high frequencies of resistance gene breakdown, agreeing with similar earlier findings for the original host (3), ‘Geneva’ (Bengtsson et al. 2000; Sandskär and Liljeroth 2005), although less so with a Belgian study where infections on this host were mostly

Table 1. (Continued from previous page)

Differential host	N° sites with class 2 scores but not higher	Frequency (%) of class 2 scores but not higher	N° sites with scores >2	Frequency (%) of sites with scores >2	Category ^a
h(0)	0	0	24	100	often
h(1)	0	0	24	100	often
h(3)	2	8	20	83	often
h(8)	3	13	21	88	often
h(10)	4	17	13	57	often
h(6)	2	8	7	29	sometimes
h(6,7)	1	4	6	26	sometimes
h(13)	4	19	6	29	sometimes
h(2)	4	17	7	30	sometimes
h(4)	1	4	6	26	sometimes
h(9)	9	39	5	22	sometimes
h(14)	7	30	3	13	rarely
h(5)	2	9	1	4	rarely
h(12)	1	4	1	4	rarely
h(11)	0	0	0	0	not overcome
h(15)	0	0	0	0	not overcome

restricted to fruit (Lefrancq and Lateur 2009). Therefore, these two genes (together with *Rvi1*) are de facto of low interest for breeding cultivars to be deployed in regions covered by the current VIN-QUEST network, until proof is provided that they do have a role to play in scab gene pyramids. The shift of the other four genes may lead to different conclusions about their value in plant breeding. For *Rvi10* and *Rvi13*, frequency of resistance gene breakdown calls was reduced by between 60 and 75%, and by between 25 and 30% for *Rvi6* and *Rvi7*. This indicates a relatively high frequency of class 2 calls for *Rvi10* and *Rvi13*, but fewer for *Rvi6* and *Rvi7*.

During the monitoring, we observed that scab infection of host (6,7) *M. floribunda* 821 often resulted in high severity scores, in accordance with previous findings (e.g., Bengtsson et al. 2000; Roberts and Crute 1994; Sandskär and Liljeroth 2005), suggesting that this genotype does not have any additional resistances. *Rvi6* is the most frequently used resistance gene in apple breeding. Cultivars carrying this resistance have been planted in limited numbers in many European countries for more than 40 years. Unfortunately, *avrRvi6* isolates are known to be present in multiple regions of Europe (Bengtsson et al. 2000; Lefrancq and Lateur 2009; Parisi et al. 2004), and over time they have recombined and so possibly have acquired additional virulences, enlarging the range of genotypes that they are able to infect (Caffier et al. 2015).

The change of threshold from class 2 to class 3 resulted in considerable reductions of breakdown calls for the *Rvi* genes in the category “rarely overcome” (i.e., *Rvi2*, *Rvi4*, *Rvi5*, *Rvi9*, *Rvi12*, and *Rvi14*), ranging from 60 to 80%. This indicated a high frequency of class 2 calls for all the corresponding differential hosts. If these calls really are compatible interactions, specific virulences are assumed to be present, but the corresponding isolates are not very aggressive and cannot yet effectuate resistance erosion. It could also be that the acquisition of these specific virulences may have a fitness cost. However, over time they may become more aggressive through directional selection, as observed previously for scab resistances in some hosts, such as *M. floribunda*, *M. ‘Mary Potter,’ M. ‘Professor Sprenger,’ M. ‘Sentinel,’* and *M. ‘SugarTyme’* (Beckerman et al. 2009).

The monitoring of resistance gene breakdowns in multiple sites provides the most comprehensive, nevertheless limited, picture of to what extent and where the different *Rvi* genes are currently overcome. The results of this analysis can be used by apple breeders in selecting gene combinations conferring more durable resistance. The limited breeding value of the *Rvi1*, *Rvi3*, and *Rvi8* genes is confirmed by the very high frequency of sites with resistance breakdown (83 to 100%). Possibly, *Rvi10* could also be added to this group. Although this gene is rarely overcome considering its overall low

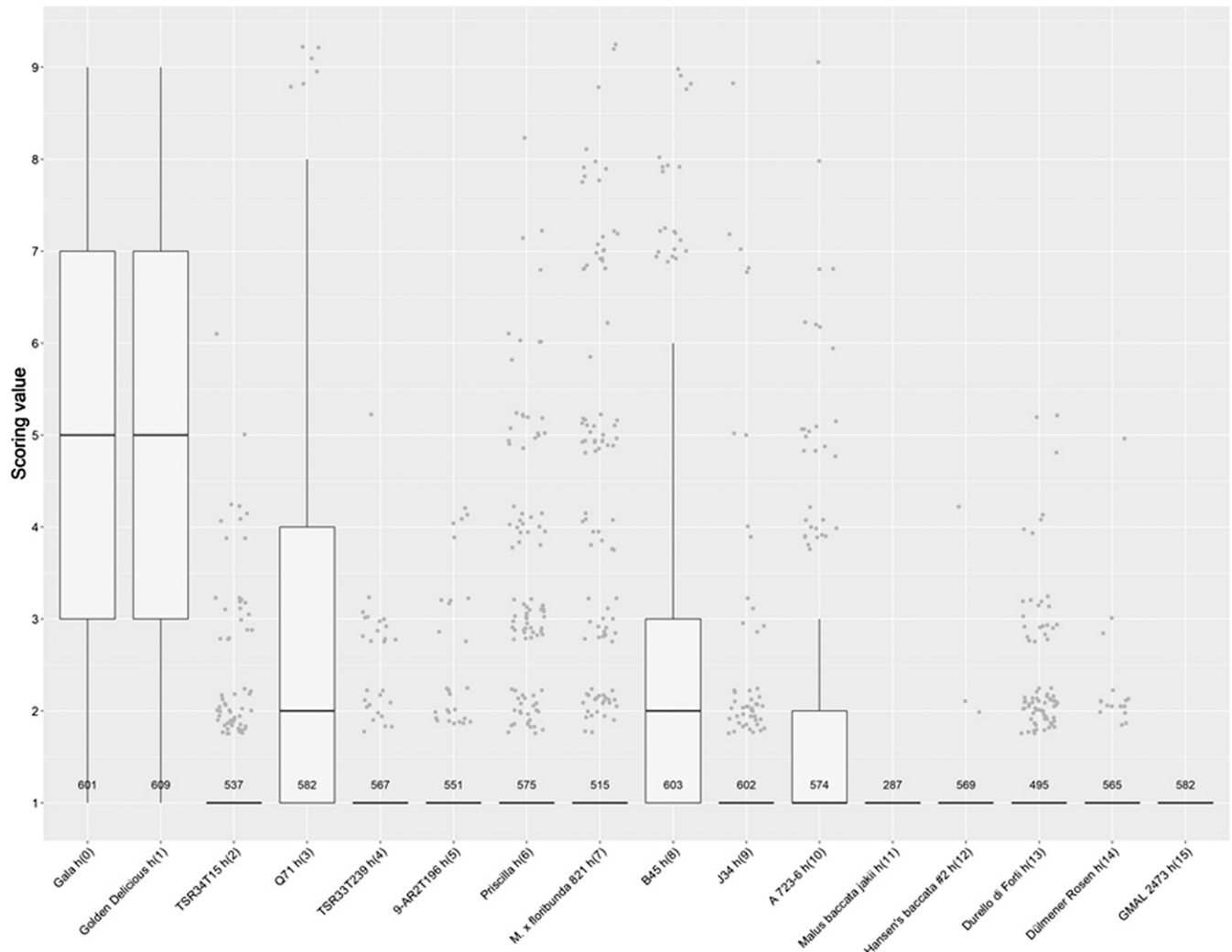


Fig. 1. Box plot of the scores observed over all years and sites for each of the 16 differential hosts. Severity class 1: No visible lesions; class 2: one or very few lesions detected upon close scrutiny of the tree ($\pm 1\%$ of leaves affected); class 3: immediately apparent lesions in general clustered in a few parts of the tree (1 to 5% of leaves affected); class 4: intermediate; class 5: numerous lesions widespread over a large part of the tree ($\pm 25\%$); class 6: intermediate; class 7: severe infection with half of the leaves badly infected by multiple lesions ($\pm 50\%$); class 8: intermediate ($\pm 75\%$); class 9: tree completely affected with (nearly) all the leaves badly infected by multiple lesions ($>90\%$). The bold horizontal line represents the median value. The boxes cover 50% of the middle values while the whiskers each cover $\sim 25\%$ of the more divergent values. The dots (outliers) are defined as values located at a distance from the median that is greater than two times the standard deviation.

frequency of scores >2, the scorings emanated from more than half of the monitored sites (13 out of 23 sites). However, in four of the sites, there was only one score >2 (Fig. 2), and it may be too soon to give more definite advice on its use in breeding.

Applying the same criteria, the category of “rarely overcome genes” based on the total number of scores >2 could be further divided into two categories: those “sometimes overcome” (*Rvi2*, *Rvi4*, *Rvi6*, *Rvi7*, *Rvi9*, and *Rvi13*) and those rarely overcome (*Rvi5*, *Rvi12*, and *Rvi14*) over the sites. In the first group we find *Rvi2*, *Rvi4*, and *Rvi6*, which are probably the most commonly used scab resistance genes in apple breeding. *Rvi2* and *Rvi4* are combined in the cultivar Regia, whose resistance is already overcome (Peil et al. 2018), and elite parents with all three genes in homozygous state have been generated (Baumgartner et al. 2015). Because of the relative high frequency (about one third) of trap orchards with isolates that were virulent to these genes, we advise that they (including also *Rvi9* and *Rvi13*) are used only in pyramids of at least three *Rvis*. As isolates combining *avrRvi6* and *avrRvi7* have been found (all those that generated infections on h(6,7)), pyramids with these two genes should be avoided or complemented with at least two additional *Rvi* genes.

Resistance gene breakdown of *Rvi5*, *Rvi12*, and *Rvi14* has rarely been encountered to date. These genes, together with *Rvi11* and *Rvi15*, which so far have not been overcome, currently constitute the best sources of apple scab resistance, thus showing most promise for durable resistance. These findings agree with host (11) *M. baccata* ‘Jackii,’ as well as many other accessions, remaining scab-free or very rarely becoming infected in long-term disease monitoring projects (Beckerman et al. 2009; Den Boer and Green 1995). Nevertheless, at times these differential hosts do get infected to some extent (Bengtsson et al. 2000; Lefrancq and Lateur 2009; Sandskär and Liljeroth 2005), but probably only when infection conditions are optimal. In 2011, limited sporulation accompanied by chlorosis was observed on host (11) *M. baccata* ‘Jackii’ in Poland (Fig. 3). Reinoculation of h(11) with the strain that was isolated from an affected leaf showed the same symptoms of partial resistance, i.e., chlorosis with limited sporulation. Hence, the initial score of 3 was corrected to 1.

Further research on the genetics of the differential hosts is required since some of these accessions, e.g., *M. baccata* ‘Jackii’ (h(11)),

Hansen’s *baccata* #2 (h(12)), and ‘Dülmener Rosenapfel’ (h(14)), carry more than one scab resistance gene (Soufflet-Freslon et al. 2008; Bus and Patocchi, unpublished data). Scoring of disease symptoms in this study may therefore underestimate infection frequencies of some genes, overestimating their breeding value. Further genetic analyses are in progress on these accessions in order to select single-gene differential hosts for *V. inaequalis* virulence monitoring in trap orchards.

The VINQUEST initiative has started well and the network continues to expand. To date, most of the collected data were contributed by European partners, while 15 partners in an additional nine countries are in the process of establishing their trap orchards. Finally, nine more partners in a further three countries have declared an interest in joining the initiative, but for different reasons have not yet done



Fig. 3. Apple scab lesions on h(11) *Malus baccata* ‘Jackii’ found in 2011 at Skierniewice (Poland). Although sporulation is obvious, this is accompanied by a resistance reaction (chlorosis) of the tree. The same reaction was obtained by artificial inoculation (data not shown). Such symptoms are not considered as a compatible interaction (abundant sporulation without reaction of the plant), and symptoms on this tree should therefore be called as severity score 1.

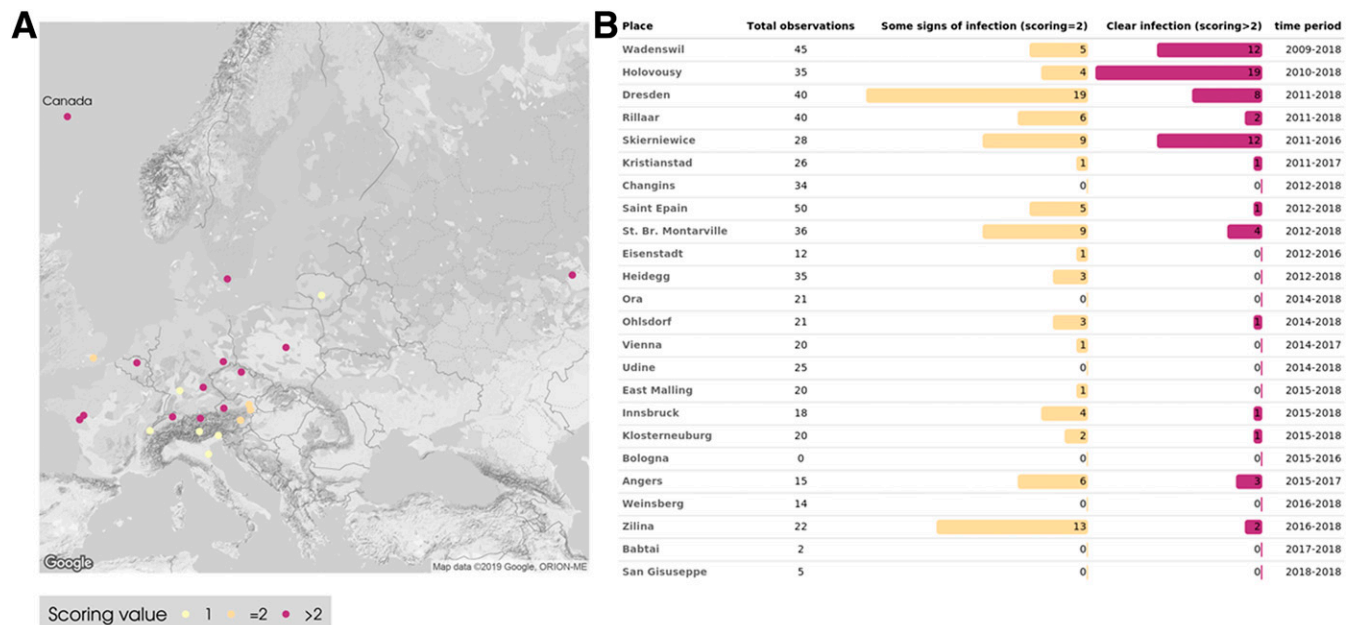


Fig. 2. A, Resistance gene breakdown of *Rvi10*. Dots represent the geographic distribution of the VINQUEST partners providing data between 2009 and 2018. For graphical reasons, Canada has been moved closer to Europe. Yellow dots: sites without observations of resistance gene breakdown. Orange dots: sites with observations never higher than severity score 2. Magenta dots: sites with at least one observation higher than severity score 2. **B,** Summary of the observations of *Rvi10* resistance gene breakdown with severity scores = 2 or >2 and indication of the periods when these observations were made.

so. Partners in South and North America are awaiting the release of plant material from quarantine, while partners in Asia are at the orchard establishment phase. Therefore, a much more comprehensive global picture of the distribution of different virulences in *V. inaequalis* populations will become available in the future.

The results of the first 10 years of VINQUEST provide the first indications on which apple scab resistance genes show the most promise for breeding. However, since most of these genes do not occur in any commercial cultivars, they have not yet been exposed extensively to scab populations at large. Therefore, all the conclusions made for the different *Rvis* should be considered as best-case scenarios, hence the deployment of resistance genes in pyramids only remains imperative to improve the chances of their durability. Of the five genes showing the most promise of achieving this, *Rvi5* and *Rvi14* are present in 'Murray' and 'Dülmener Rosenapfel' (recognizing that this cultivar carries multiple resistances), respectively, while *Rvi15* is present in GMAL2473, a genotype producing relatively large apples. However, they all need further improvement for eating quality to meet today's breeder and consumer expectations. Nevertheless, they are at an advantage compared with the *Rvi11* and *Rvi12* genes, which are present in small-fruited wild apples, and so are expected to require more generations of backcrossing to improve both fruit size and quality. Some breeding programs have already performed several backcross cycles, but those still in the early stages would greatly benefit from using fast-breeding techniques, such as Fast-track (Volz et al. 2009) and the early flowering approach using the *BpMADS4* gene (Flachowsky et al. 2011; Schlathölter et al. 2018) to accelerate the removal of undesired traits present in the donors of these two resistances. While not considered to produce a genetically modified plant in some countries, the application of this later new breeding technology defines it as a genetically modified plant in others (Bus et al. 2019).

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